Studies in *Oberonia* 10. A re-evaluation of *Oberonia brachystachys* and *O. subligaculifera* (Orchidaceae: Malaxideae) and their synonyms

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Abstract: The following synonyms are introduced or confirmed for Oberonia brachystachys Lindl.: O. recurva Lindl., O. setifera Lindl., O. demissa Lindl., O. gardneriana Thwaites, O. parvula King & Pantl. [including O. myriantha var. parvula (King & Pantl.) Tang & Wang], O. croftiana King & Pantl. (second-step lectotypification herein), O. lingmalensis Blatt. & McCann [including O. recurva var. lingmalensis (Blatt. & McCann) Santapau & Kapadia], and O. brachyphylla Blatt. & McCann (lectotypified herein). Oberonia subligaculifera J.J.Sm. has the following new or confirmed synonyms: O. bambusicola Kerr, O. evrardii Gagnep., O. kanburiensis Seidenf., O. arunachalensis A.N.Rao, O. raoi L.R.Shakya & R.P.Chaudhary, and O. kamlangensis A.N.Rao. Both species are re-described taking into account their intraspecific variability and ecological data are provided. Oberonia brachystachys with unjointed leaves is illustrated with light and scanning electron microscope images.

Keywords: Lectotypification, Morphological characters, Nomenclature, Variability.

Introduction

Oberonia Lindl. is an old-world genus of mostly epiphytic herbs comprising 300 species (Mabberley, 2018), which is likely an overestimate given the numerous new synonyms uncovered in recent years (Geiger, 2016, 2019, 2020, 2021, unpubl. data; Gogoi & Yonzone, 2016; Bunpha *et al.*, 2019; Geiger *et al.*, 2020). Geiger *et al.* (2020) estimated

Received: 23.11.2020; *Revised & Accepted*: 16.09.2021 *Published Online*: 16.10.2021 that the genus is over-described by approximately one third, which would put the true species number rather at approximately 150–200 worldwide (Pridgeon *et al.*, 2005).

In India, 61 species have been reported in the genus (Ansari & Balakrishnan, 1990; Kumar & Manilal, 1994, 2004; Misra, 2007; Narayanan et al., 2010, 2013; Chowlu et al., 2015; Chowlu & Rab, 2017; Prasad et al., 2018; Kaliamoorthy & Saravanan, 2019). That number of species is equally an overestimate due to a number of synonyms uncovered in recent years (Geiger, 2016, 2019, 2020; Gogoi & Yonzone, 2016; Bunpha et al., 2019): O. anthropophora Lindl. (+ O. falcata King & Pantl.), O. bicornis Lindl. (+ O. tenuis Lindl.), O. brunoniana Wight (+ O. lindleyana Wight, O. saintberchmansii Kad.V.George & J.Mathew, O. santapaui Kapadia in Santapau & Kapadia, O. wallichiii Hook.f.), O. caulescens Lindl. (+ O. katakiana A.N.Rao), O. langbianensis Gagnep. (+ O. sulcata Joseph & Sud.Chowdhury), O. maxima Parish ex Hook.f. (+ O. integerrima Guillaumin, O. orbicularis Hook.f.), O. mucronata (D.Don) Ormerod & Seidenf. (+ O. manipurensis Chowlu, Y.N.Devi, A.N.Rao, N.Angela, H.B.Sharma & Akimpou), O. rufilabris Lindl. (+ O. nepalensis L.R.Shakya & R.P.Chaudhary, O. pantlingiana L.R.Shakya & R.P.Chaudhary). It is likely that additional names will be placed in synonymy.

A species of *Oberonia* collected during botanical explorations in Kerala (2019–2020), in the Aralam

Wildlife Sanctuary, Kannur, flowered in the Glass House at the Botanical Survey of India, Southern Regional Centre, Coimbatore. It was identified as *O. brachystachys* Lindl. based on herbarium records at CAL and MH, and literature (Seidenfaden, 1968; Ansari & Balakrishnan, 1990; Misra, 2007). The specimen was initially considered a range extension of *O. brachystachys*, but provided the impetus for a critical taxonomic analysis resulting in the unravelling of the convoluted taxonomy.

Materials and Methods

Taxonomic assessments were made based on available information from primary and secondary literature, herbarium holdings (BM, CAL, E, F, K, MEL, MO, OHT [Orchid Herbarium, Tipi, Arunachal Pradesh, India], P, US, W, WU), and on-line herbarium databases (AMES, BR, L; herbarium acronyms as per Thiers, continuously updated). The K specimens without barcode are held in the Lindley herbarium at K. DLG numbers refer to the Daniel L. Geiger living collection in greenhouse in Santa Barbara, California. HOAG numbers refer to the Geiger personal herbarium.

The type designation follows the modern interpretation of McNeill (2014, 2015). Unless a holotype is designated explicitly with repository in the protologue, or the description is explicitly based on a single specimen, then all specimens from the cited gathering(s) are syntypes.

Ecological data was gathered from the literature and indications on herbarium specimens, with all herbariums specimens being considered equivalent to one literature reference. The percentage of observations refer to the frequency a particular flowering month or elevation (to nearest 100 m) has been given by each of the cited sources. Accordingly, 100% means that all cited sources agree, while 50% indicates that only half of the sources give that particular flowering month or elevation.

Z-stack photography was carried out on a Discovery V20 stereomicroscope with plan

apochromatic lenses and motorized focus (Zeiss, Germany). Image stacks were captured with an Axiocam HRc camera (Zeiss, Germany) and processed in ZereneStacker (Zerene Systems LLC, 2009–2021). Flowers and floral parts were also imaged with a SMZ1500 stereomicroscope (Nikon, Japan). For scanning electron microscopy (SEM), flowers were preserved in 95% ethanol, further dehydrated through three changes of 100% ethanol, critical point dried in a 815A (Tousimis, USA) using standard settings, then mounted on double sided carbon tabs (Ted Pella, USA), sputter coated with gold, and imaged in an EVO 40 XVP (Zeiss, USA) using the VPSE detector in variable pressure (30 Pa) at 20 kV and probe currents ranging from 30-500 pA depending upon working distance and magnification. Illustrations were processed in AffinityPhoto 1.10.1 (Serif, 2021).

Taxonomic Treatment

Oberonia brachystachys Lindl., Sert. Orchid. t. 8 B. 1838. *Syntypes*: BURMESE EMPIRE, *s.loc., s.d., Griffith* 697 (K!), *Griffith* 778 (K!; P [P00404973]!); possible syntype: *Griffith s.n.* (K!).

Oberonia recurva Lindl., Edwards's Bot. Reg. 25 (Misc.): 14. 1839. *Lectotype* (inadvertent designation by Ansari & Balakrishnan, 1990: 38 by reference to "Type ... (K)"): INDIA, **Maharashtra**, Bombay, *s.d., Loddiges s.n.* (K!), **syn. nov.**

Oberonia setifera Lindl., Fol. Orchid. 8: 3. 1859. *Lectotype* (inadvertent designation by Ansari & Balakrishnan, 1990: 38 by reference to "Type ... (K)"): INDIA, **Maharashtra**, South Koncan (= Koukan, Mumbai), *s.d., Dalzell* ex *Stocks* 38 (K!), **syn. nov.**

Oberonia demissa Lindl., Fol. Orchid. 8: 4. 1859. *Lectotype* (inadvertent designation by Seidenfaden, 1978: 37 by reference to "K, type"): INDIA, **Sikkim**, Terai, *J.D. Hooker* 121 (K!).

Oberonia gardneriana Thwaites, Enum. Pl. Zeyl. 296. 1861. *Syntype:* SRI LANKA, Ambagamowa district, *s.d., Gardner s.n.* [*Thwaites*] *CP* 593 (PDA *n.v.* fide Jayaweera, 1981: see Notes). **syn. nov.** *Oberonia parvula* King & Pantl., J. Asiat. Soc. Bengal, Pt. 2, Nat. Hist. 64: 330. 1896. *Lectotype* (inadvertent designation by Ansari & Balakrishnan, 1990: by reference to "Type: *Pantling 203* ... (CAL)"]: INDIA, **Sikkim**, valley of Teesta, 1500 ft, *R. Pantling* 203 (CAL [CAL000000040]!); *isolecto* AMES/HUH [HUH00102050]!, BM [BM000088303]!, G *n.v.*: fide Seidenfaden, 1968, K [K000387743]!, P [P0040885]!). **syn. nov.**

Oberonia croftiana King & Pantl., Ann. Roy. Bot. Gard. (Calcutta) 8: 7, pl. 6A. 1898. *Lectotype* (firststep inadvertent designation by Ansari & Balakrishnan, 1990: 38 by reference to "Type: ...(CAL)"; second-step designation here): INDIA, **Sikkim**, Tropical Valleys, October 1893, *Pantling* 254 (CAL [CAL0000000048: Tropical Valleys]!), *isolectotypes* (CAL [CAL0000000049]!; AMES [HUH 00101950]!, BM [BM000088297]!, BR [BR0000006572181]!, K [K000387742]!, P [P00404879]!). *Residual syntypes*: INDIA, **Sikkim**, Dikkeling [= Dikling], *Pantling* 254 L [L0061772]!; W [W5951]!); Locality unknown (G *n.v.*: fide Seidenfaden, 1968). **syn. nov.**

Oberonia lingmalensis Blatter & McCann, J. Bombay Nat. Hist. Soc. 35: 255. 1931. O. recurva Lindl. var. lingmalensis (Blatt. & McCann) Santapau & Kapadia, J. Bombay Nat. Hist. Soc. 57: 259. 1960. Holotype: INDIA, Maharashtra, Western Ghats, Lingmala, near Yenna waterfall, a few miles from Mahableshwar [= Mahabaleshwar] on road to Panchgani, Blatter & Hallberg P 1681 (BLAT *n.v.*). Paratypes: INDIA, Karnataka, North Kanara, in forests, Bell 5406 (repository unknown). Maharashtra, Kamelgad, below Fort (Fernandez I). Sedgwick 7755, 4626 (repository unknown). Maharashtra, Konkan, Thana [= Thane] forest, Bell 3972 (repository unknown). syn. nov.

Oberonia brachyphylla Blatt. & McCann, J. Bombay Nat. Hist. Soc. 35: 257. 1931. *Lectotype* (designated here): INDIA, **Karnataka**, North Kanara, *Blatter* & *McCann* (1931: pl. 2!). **syn. nov.** Figs. 1–5

Small erect or pendulous epiphytic herbs, usually 4–6 leaves, unjointed, forming stemless fan. Leaves

variable in length, shape, typically elongated ovate acute, occasionally squat ovate acute, typically 2-6 cm long, 0.6–1.2 cm wide. Roots thin, fibrous, unbranched. Inflorescence typically 5–7 cm long, typically 1.5–3× as long as leaves. Scape short, rachis longitudinally grooved, flowers variously arranged from distinct whorls, to in spiral, or scattered. Flowers 0.75–1.5 × 1–1.75 mm, average ~1.5 × 1.5 mm, green, yellow, brown, orange, sometimes two-coloured with lip orange to brownish red, reminder greenish. Bract acuminate acute, margin irregular, about as long as flower. Pedicelled ovary round, ~1 mm long. Orientation of all tepals flat or somewhat forward. Sepals broad, oval, acute. Petals oblong, obtuse, erose to dentate. Lip typically four lobed, lateral lobes square or trapezoidal with tips towards epichile, margin typically erose to dentate, occasionally irregular; mesochile more or less constricted, entire; typically two distinct (occasionally indistinct) epichile lobes, truncated, rounded or falcate towards midline, margins dentate, erose, or occasionally irregular, disc indistinct, sac minute depression to large cavity.

Flowering: Flowering throughout the year with peaks in February and October (Fig. 5a). The break in June is considered a data artifact.

Habitat: Erect or pendulous epiphyte on branches of shrubs and trees in tropical wet or dry evergreen forest to subtropical montane forest, in association with Dendrobium ovatum (L.) Kraenzl. (Orchidaceae), on Carissa congesta L. (Apocynaceae), Anacardium occidentale L., Holigarna grahamii (Wight) Kurz, Mangifera indica L. (all Anacardiaceae), Heterophragma quadriloculare (Roxb.) K.Schum. (Bignoniaceae), Garcinia indica Choisy (Clusiaceae), Elaeocarpus serratus L. (Eleocarpaceae), Euphorbia neriifolia L. (Euphorbiaceae), Erythrina suberosa Roxb., Bauhinia racemosa Lam. (all Fabaceae), Memecylon umbellatum Burm.f. (Melastomataceae), Tectona grandis L.f. (Lamiaceae), *Careya arborea* Roxb. (Lecythidaceae), Bombax ceiba L. (Malvaceae), Ficus arnottiana (Miq.) Miq., F. benghalensis L., F. racemosa L., Artocarpus heterophyllus Lam. (all Moraceae),

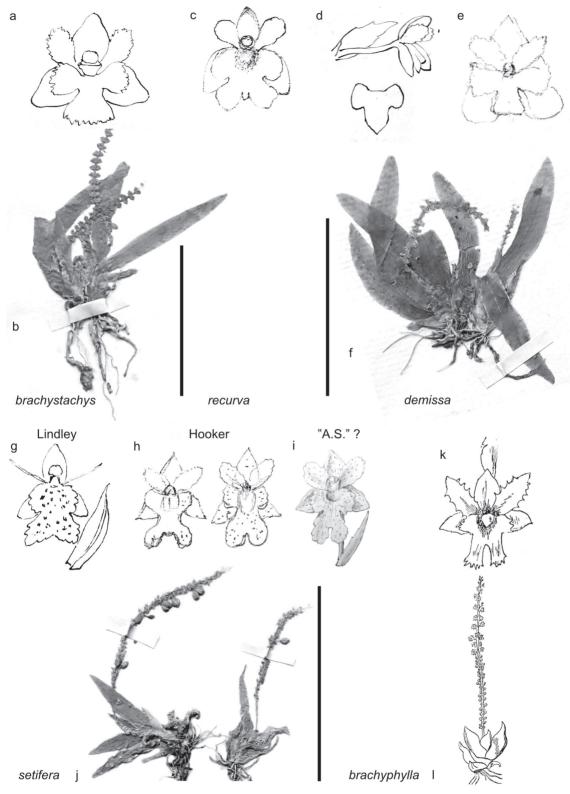


Fig. 1. Photographic images and drawings of *Oberonia* spp.: a & b. *O. brachystachys* Lindl., K syntype sheet; the two gatherings *Griffith* 697 and *Griffith* 778 are not individually identified on sheet; a. Drawing of flower; b. Habit of one of three specimens; c. *Oberonia recurva* Lindl., drawing of flower on lectotype sheet *Loddiges* s.n (K); d–f. *Oberonia demissa* Lindl., *Hooker* f. 121 (lectotype K); d. Drawing by unknown person; e. Drawing by J.D. Hooker; f. Specimen; g–j. *Oberonia setifera* Lindl., *Dalzell* ex Stocks 38 (lectotype K); g. Drawing by J. Lindley; h. Drawings by J.D. Hooker; i. Drawing by "A.S." [?]; j. Specimen; k & I. Lectotype of *O. brachyphylla* Blatter & McCann (1931: pl. 2); k. Flower; I. Habit. Scale bars b, f, j = 5 cm. (photos a–j by D. L. Geiger; k, I from Blatter & McCann [1931: pl. 2]).

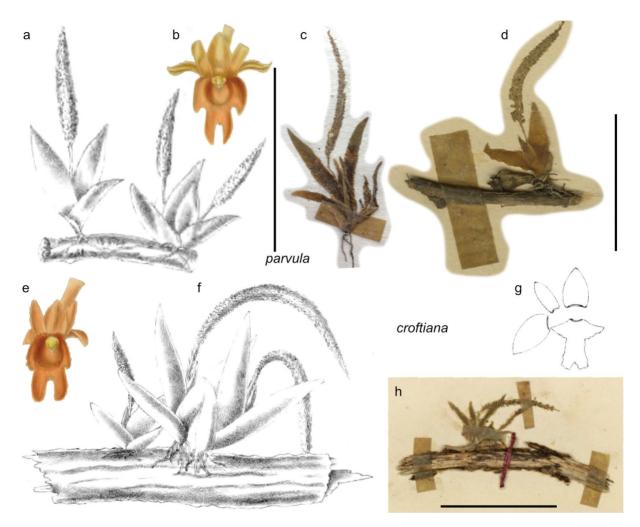


Fig. 2. Photographic images and drawings of *Oberonia* spp. a. Drawing of habit of *O. parvula* King & Pantl. from King and Pantling (1898: pl. 6B); b. Flower of *O. parvula* from King and Pantling (1898: pl. 6B); c. Likely original material of *O. parvula*, *Pantling* 203 (K [K000387743]); d. Likely original material of *O. parvula*, *Pantling* 203 (AMES [HUH00102050]); e. Flower of *O. croftiana* King & Pantl. from King and Pantling (1898: pl. 6A); f. Drawing of habit of *O. croftiana* from King and Pantling (1898: pl. 6A); g. Drawing of flower on *O. croftiana* isolectotype sheet *Pantling* 254 (P [P00404879]). Floral parts digitally rearranged to natural positions; h. Isolectotype *Pantling* 254 (L [L0061772]). Scale bars c,h = 5 cm; d = 2.5 cm. (a,b,d–h from cited publication under fair use provision of copyright law. Photo c by D.L. Geiger).

Canthium dicoccum Gaertn., Catunaregam spinosa (Thunb.) Tirveng., Ixora brachiata Roxb., Randia brandisii Gamble (all Rubiaceae), Flacourtia indica (Brum.f.) Merr. (Salicaceae), Turpinia malabarica Gamble (Staphyleaceae). 0–2400 m, predominantly below 1800 m (Fig. 5b).

Distribution: Bhutan, China (Guangxi?: see Notes), India, Malaya, Myanmar, Nepal, Sri Lanka, Thailand (Anonymous, n.d.b; Hooker, 1888; Cooke, 1907; Brühl, 1926; Blatter & McCann, 1931; Mitra, 1958; Seidenfaden & Smitinand, 1959; Santapau, 1967; Seidenfaden, 1978; Pradhan, 1979; Bose & Bhattacherjii, 1980; Jayaweera, 1981; Banerji & Pradhan, 1984; Ansari & Balakrishnan, 1990; Naithani, 1990; Seidenfaden & Wood, 1992; Turner, 1995; Sakkar, 1995b; Srivastava, 1996; Bose *et al.*, 1999; Kress *et al.*, 2003; Lucksom, 2007; Misra, 2007; Raskoti, 2009; Chen *et al.*, 2009 Cerejo-Shivkar & Shinde, 2015; Jalal, 2018; Singh *et al.*, 2019: see Notes). Some of the eastern distributional records (Malaya, Thailand) may be due to confusion of *O. brachystachys* with *O. subligaculifera*. The distribution based on type material of the *O. brachystachys* and its synonyms extends from India, Sri Lanka, to Myanmar.

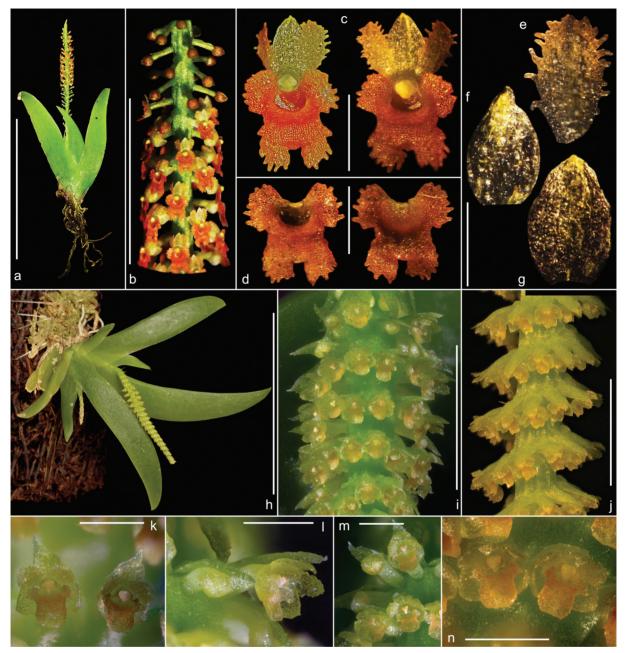


Fig. 3. Photographic images of living plants of *Oberonia brachystachys* Lindl.: **a.** Habit; **b.** Portion of inflorescence; **c.** Flowers; **d.** Lip, front (left) and back (right) sides; **e.** Petal; **f.** Lateral sepal; **g.** Median sepal; **h.** Habit (DLG521); **i.** Portion of inflorescence, z-stack of 42 images (DLG521); **j.** Portion of inflorescence, z-stack of 10 images (DLG521); **k.** An. Individual flowers (DLG521): **k.** Z-stack of 43 images (DLG522); **I.** Z-stack of 38 images; **m.** Z-stack of 36 images; **n.** Z-stack of 53 images. Scale bars: a,h = 5 cm; b = 1 cm; c,d,k–n = 1 mm; e–g = 0.5 mm; i,j = 5 mm (photos a–g by M. Sulaiman, from *M. Sulaiman* 145464, MH [MH00257633]; h–n by D. L. Geiger).

Specimens examined: INDIA, Kerala, Kannur, Aralam Wildlife Sanctuary, N 11°552 512 2 , E 75°482 452 2 ± 110 m, 29.12.2019, *M. Sulaiman* 145464 MH [MH00257633]. Maharastra, Western Ghats, Lingmala near Mahabaleshwar, *Townsend* 73/10 (K). Sikkim, valley of theTeesta, 1898, *R. Pantling* 238 (BM [BM000088265], CAL

[CAL0000077889], MEL [MEL2409511A], MH [MH00042702], P [P00404975], P[P00404976], US [US00241398], WU).

Notes: *Oberonia brachystachys* is characterized by the relatively small size of the plant, unjointed leaves, bracts that are about as long as the pedicellate ovary,

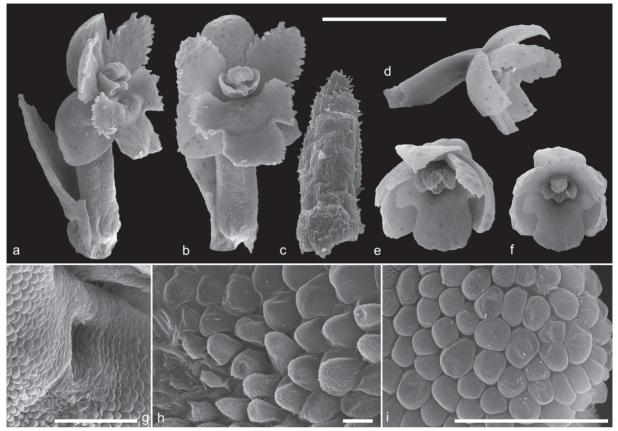


Fig. 4. SEM images of *Oberonia brachystachys* Lindl.: **a.** Flower–oblique view with floral bract; **b,e & f.** Flower–frontal view; **c.** Floral bract; **d.** Flower–lateral view; **g.** Disc with sac; **h.** Microrugulate surface morphology of conical cells surrounding disc; **i.** Microrugulate surface sculpture of pneumate epichile cells. Scale bar for a-f = 1 mm; $g,i = 100 \text{ }\mu\text{m}$; $h = 10 \text{ }\mu\text{m}$; (a-c,g-i, K [K59697], India; d-f. DLG521/HOAG78, in cultivation; images by D.L. Geiger).

small flowers ($0.75-1.5 \times 1.0-1.75$ mm according to protologues, average ~ 1.5×1.5 mm), strongly erose, ovate petals, and a variable lip typically with quadrate lateral lobes with strongly erose margins, a more or less constricted and entire mesochile, and more or less developed diverging epichile lobes with strongly erose margins.

Oberonia brachystachys has two recognized synonyms, *O. demissa* and *O. bambusicola* Kerr (Seidenfaden & Smitinand, 1964; Bunpha *et al.*, 2019). *Oberonia demissa* is here confirmed as synonym of *O. brachystachys*, while *O. bambusicola* is synonymized under *O. subligaculifera* (see below). Below we discuss each of the synonyms in greater detail.

Three main forms have been recognized, distinguished by the unnotched epichile of *O. brachystachys vs.* deeply incised epichile and more

or less rectangular epichile lobes of *O. recurva vs.* lip with falcate epichile lobes of *O. parvula*. Those three morphs represent points along a spectrum of continuous variability. The three lips illustrated by Ansari and Balakrishnan (1990: fig. 36) under *O. recurva* show variation from shallow notch to deep incision, clearly documenting the intraspecific variability. Illustrations of *O. brachystachys* (Fig. 1) and its synonyms *O. parvula*, *O. croftiana* (Fig. 2) show intermediate character states of the epichile lobes quadrate, rounded, and falcate.

Differences in drawings from the same plant as demonstrated here with *O. setifera* (Fig. 1g–i) further caution the reliance on minute details in drawings. Floral variability has been confirmed with molecular data in *O. equitans* (G.Forst.) Mutel by Geiger *et al.* (2020) is also evident in the work of Li *et al.* (2016) (see Geiger *et al.*, 2020 for details).

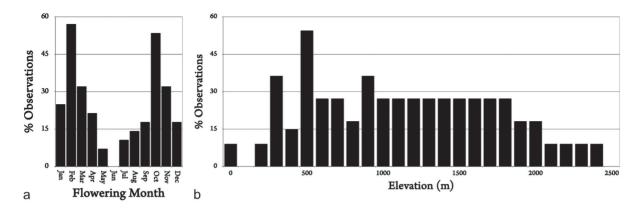


Fig. 5. Summary bar charts of ecological data for *Oberonia brachystachys* Lindl., compiled from literature under any of its synonyms: **a**. Phenology data from Ansari and Balakrishnan (1990), Banerji and Pradhan (1984), Blatter and McCann (1931), Bose and Bhatacherjii (1980), Bose *et al.* (1999), Brühl (1926), Bunpha *et al.* (2019), Cooke (1907), Cerejo-Shivkar and Shinde (2015), Lucksom (2007), Jalal (2018), Jayaweera (1981), King and Pantling (1898), Nayar *et al.* (2006), Pradhan (1979), Rao (2000), Raskoti (2009), Santapau (1953, 1967), Santapau and Kapadia (1960), Srivastava (1996), herbarium records, Geiger (pers. obs. *in cult.*); **b**. Elevation data from Ansari and Balakrishnan (1990), Bose *et al.* (1999), Brühl (1926), Bunpha *et al.* (2019), De Vlas (2019), Jalal (2018), Jayaweera (1981), King & Pantling (1898), Lucksom (2007), Mitra (1958), Rajbhandari (2015), Raskoti (2009), Kumar and Manilal (1994), Seidenfaden (1978), Seidenfaden and Smitinand (1959), Srivastava (1996); (graphics by D.L. Geiger).

Oberonia recurva (Fig. 1c) is a synonym of *O. brachystachys.* Both species were introduced by Lindley (1838, 1839). *Oberonia recurva* was described with erose petals (*petalis obovatis subdentatis*), and a four-lobed lip (*labello subrotundo quadrilobo denticulato mucrone interjecto*). It was only compared to *O. wightiana* Lindl., with a much larger flower, linear almost entire petals, and with lateral lobes folded over the midline of the flower. It is a typical case where a new species is justified by comparison to a very dissimilar species, as opposed to the most similar ones. The lectotype at K agrees with the protologue.

While several names have been considered synonyms of O. *recurva* (O. *setifera*, O. *gardneriana*, O. *parvula*, O. *croftiana*, O. *lingmalensis*, O. *myriantha* var. *parvula*: Jayaweera, 1981; Ansari & Balakrishnan, 1990; WCSP, 2020), those have not been discussed and justified and are treated here in more detail.

Oberonia setifera Lindl. (Fig. 1g–j; not to be confused with *O. setigera* Ames from the Philippines) was described with awn-shaped petals (*petalia setaceis*), and a four-lobed, denticulated lip, and its small vegetative size was noted. The flowers were referred to as being perhaps the smallest in the genus. The lectotype at K agrees with *O. brachystachys*. The description of the petals is odd and may be the result of folded over petals.

The drawings on the type sheet of O. setifera by three different authors are illuminating, as it demonstrates the range of interpretations based on the same specimen. Lindley's drawing (Fig. 1g) has rather narrow petals as noted in the protologue and the four lobes of the lip are rather different from the other drawings, but show an erose margin of the lip. The drawings by Hooker (Fig. 1h) and A.S. [? handwriting difficult to read] (Fig. 1i) show the erose and wider petals, and a more typical fourlobed lip. The drawings by Hooker do not show the erose margin of the lip, while that of A.S. illustrates it very clearly. Either those differences are due to artistic license or due to minute differences of flowers from the same plant. Regardless, it shows that those difference are meaningless at the species level and are either variability or artistic license.

Oberonia demissa (Fig. 1d–f) is a synonym of *O. brachystachys*. The small plant with small flowers has unjointed leaves and strongly erose petals. The lip was described by Lindley (1859) as trifid with ovate acute middle lobe [= epichile]. That condition is shown in a drawing (Fig. 1d: presumably by

Lindley) on the lectotype at K. A second drawing of the flower by J.D. Hooker (Fig. 1e), who collected the plant, shows the more typical fourlobed lip of *O. brachystachys*. The lobes are shown not as large and distinct as is typical, which could either be an inaccuracy of the drawing or a limitation of the preservation. The state of the specimen did not permit to clearly see the condition of the epichile. The combination of plant size, unjointed leaves, and deeply erose petals is consistent with the identification of this plant as *O. brachystachys*. The drawings of the flowers of *O. brachystachys* (Fig. 1a) and *O. demissa* by Hooker (Fig. 1e) are indistinguishable.

The exquisite drawing under *O. demissa* by King and Pantling (1898: pl. 10) shows a flower with lateral lobes of the lip drawn downwards, a distinct constriction of the mesochile, and very small epichile lobes. It appears intermediate between *O. brachystachys* with sub-quadratic lateral lobes and distinct constriction of the mesochile, and the *O. parvula*-morph (see below) with drawn down epichile lobes but little constriction of the mesochile. It is not clear which plant formed the basis of this drawing and, hence, the accuracy of the drawing could not be verified.

Oberonia gardneriana is a synonym of O. brachystachys. It was described with a bract more or less as long as the pedicelled ovary, oblong, blunt and denticulate petals (petalis oblongis, obtusis, denticulatis), and a red lip. The lip was not further described and no size indications were provided. It was compared only to O. wightiana (see above for differentiation). While the description is far from complete, that combination of characters is most compatible with O. brachystachys amongst the species known from Sri Lanka (Jayaweera, 1981; Fernando et al., 2003; de Vlas, 2019). Members of the section Scyllae (O. bicornis Lindl. = O. tenuis Lindl.: Geiger, 2019], O. claviloba Jayaw., O. dolabrata Jayaw., O. fornicata Jayaw., O. meegaskumburae Priyad., Wijew. & Kumar, O. scyllae Lindl., O. walliesilvae Jayaw., O. weragamaensis Jayaw.) have pubescent edges of the petals. This section of Oberonia is in need of critical assessment, which is, however, beyond the scope of this contribution. *Oberonia forcipata* Lind., *O. truncata* Lindl., *O. wightiana*, and *O. zeylanica* Hook.f. have linear lanceolate petals, *O. longibracteata* Lindl. has bracts that are much longer than the pedicelled ovary, *O. quadrilatera* Jayaw. has sessile flowers, and the petals of *O. thwaitesii* Hook.f. are very similar to the ovate triangular sepals. Accordingly, with the available data and historical precedents, it is most likely that *O. gardneriana* is conspecific with *O. brachystachys.* Jayaweera (1981), who had seen the type, listed *O. gardneriana* under *O. recurva*, itself a synonym of *O. brachystachys.*

The gathering of the type of *O. gardneriana* is ambiguous. Thwaites (1861) gave *Gardner*, in italics but no number, suggesting a gathering. He also gave the number C.P. 593, in roman font, a letter-number combination frequently seen with Thwaites material. As the only specimen at PDA could not be examined and PDA did not respond to inquiries, this matter remains to be fully resolved.

The typification of *O. parvula* presents some challenges. King and Pantling (1896) neither designated a particular type gathering, nor did they illustrate the species in the protologue itself. The likely original material (*Pantling* 203) from Teesta valley, 1500 ft, is not precisely from the type locality Guru-bathan, 1500 ft., specified in the protologue, which is located in the northern most Teesta valley. Despite this minor mismatch, *Pantling* 203 is clearly original material (See Art. 9.4, Turland *et al.*, 2018). Ansari and Balakrishnan (1990) inadvertently designated the CAL specimen as the lectotype (See Art. 9.3, Turland *et al.*, 2018).

The illustrations of the species in King and Pantling (1898: pl. 6B) with a drawing by R. Pantling, are excellent, certainly represent the original species concept, and clearly identify the species. Although some published drawings are highly inaccurate to misleading (*e.g.*, *O. carpina* Gilli: see Geiger, 2019), *Pantling* 203 (K000387743) matches the illustration by King and Pantling (1898) very well with the

pointed lateral lobes and the acuminate falcate epichile lobes.

Oberonia parvula (Fig. 2a-d) was described in the protologue from Bhutan [today India, West Bengal], Guru-bathan [= Gorubathan, N-most end of Tista river], 1500 ft, and no gathering was specified. The lectotype was designated based on likely original material from Teesta valley. The species is typically considered a synonym of O. recurva (Ansari & Balakrishnan, 1990; Rajbhandari, 2015; WCSP, 2020), but has been recognized as a distinct species by a number of authors (Brühl, 1926; Mitra, 1958; Bose & Bhattacherjii, 1980; Naithani, 1990; Sakkar, 1995a,b; Bose et al., 1999; Lucksom, 2007; Raskoti, 2009). The distinguishing characters between the O. parvula-morph and O. brachystachys include: petals only with irregular outline in O. parvula, distinctly to strongly erose in O. brachystachys; lateral lobes of lip with downwards expanding tips and entire margin in O. parvula, quadratic with erose lateral margin in O. brachystachys; mesochile barely constricted in O. parvula, strongly constricted in O. brachystachys; epichile lobes straight or curved towards midline with entire margin in O. parvula, expanding laterally away from midline with strongly erose margin in O. brachystachys. The very limited number of samples, the intermediate forms known e.g. as O. croftiana (see below), the illustration under O. demissa by King and Pantling (1898: pl. 10: see above), and the known extensive intraspecific variability in Oberonia species demonstrated by molecular phylogenetic studies (Li et al., 2016; Geiger et al., 2020), however, suggests that O. parvula is a mere morph of O. brachystachys.

The O. parvula-morph shares some floral characteristics with O. acaulis, such as the drawnout tip of the lateral lobe. Those similarity may have been the reason to consider it a variety of O. *myriantha* Lindl., a synonym of O. acaulis Griff.: O. *myriantha* var. parvula (King & Pantl.) Tang & Wang. Tang and Wang (1951) based their assessment purely on vegetative characters, but even the jointed leaves of O. acaulis do not agree with the unjointed leaves of *O. brachystachys.* Additionally, *O. acaulis* has much longer leaves even in small plants, the bract is serrated (not entire or slightly erose), the mesochile of the lip shows a distinct constriction, and the apical lobes diverge from the mid-line. The coloration of the flower typically shows a darker spot in the disc area not known from *O. brachystachys.* Accordingly, *O. parvula* is here synonymized under *O. brachystachys,* while *O. acaulis* is a distinct species. The same conclusion was reached by Seidenfaden (1968).

The specific epithets *myriantha* and *acaulis* were recently confused by Bunpha *et al.* (2019), along with several other misinterpretations about the taxonomy of *Oberonia* species. It is beyond the scope of this contribution to correct those problems.

The type material of O. croftiana presents some challenges. The protologue gave Sikkim-Bhutan frontier, on the banks of the Jaldakha river, 900 ft. as the type locality for Pantling 254. Pantling gatherings were not necessarily collected at a particular time, and may not have been from the exact same place, hence, do not constitute a gathering in the conventional sense: one species from a particular locality at a particular time (A. Schuiteman, pers. comm.). In the case of O. croftiana, Pantling 254 in W (W5951) and L (L0061772) were collected at Dikling at 2000 ft., some 20 km W of the border to Bhutan and approximately 50 km NW of the Jaldakha river, in February, 1897. Pantling 254 first-step lectotypes CAL000000048 and CAL000000049, and isolectotypes AMES 72805/HUH00101950, K000387742 and P00404879 were collected in Tropical Valley in October 1893. While the localities of all those specimens do not precisely match the indication in the protologue, they are considered here part of the lectotype series; it certainly is original material in the sense of Turland et al. (2018). CAL000000048 was annotated as "Type" by Ansari in 1985. However, no specific sheet of the two at CAL was explicitly referred to in any publications, the mention of CAL as type repository only qualifies as a first-step lectotypification (See

Art. 9.17, Ex. 14, Turland *et al.*, 2018). We here explicitly designate CAL0000000048 as lectotype by second-step lectotypification.

Oberonia croftiana (Fig. 2e-h) is a synonym of O. brachystachys. The species has been given as a synonym of O. recurva in the literature (Ansari & Balakrishnan, 1990; WCSP, 2020). It has been considered a correct name by some (Brühl, 1926; Mitra, 1958; Bose & Bhattacherjii, 1980; Naithani, 1990; Sakkar, 1995a; Bose et al., 1999; Lucksom, 2007). It is notable, that the authors who consider both O. parvula and O. croftiana distinct are almost identical. We have not found any publication that considers O. parvula with synonym of O. croftiana as distinct from O. brachystachys/recurva. The shared characters between O. croftiana and O. brachystachys include the lateral lobes of the lip with erose margin and distally extended tip, the constriction of the mesochile, and the epichile lobes with erose margin. Lucksom (2007: fig. 163) illustrated a drawing under the name O. parvula that shows an intermediate condition between the illustrations of O. parvula and O. croftiana, which lends support to the synonymy of the two taxa. Lucksom's (2007: fig. 162) illustration of O. croftiana shows a specimen with even more pronounced falcate epichile lobes. A drawing on the isolectotype (P00404879, Fig. 2g) shows rather different interpretation of the flower: petals entire and not erose, lateral lobes of the lip, not downward sloping, and epichile lobes short and square, not elongated sub-falcate. It is a further example of how the same plant may be interpreted differently. A drawing and annotation of the flower on isolectotype (CAL000000049) by Shakya in 1997 shows the lateral lobes expanding laterally without downturned tips and noted the epichile lobes are entire. Those differences compared to the drawings on the other isolectotypes and the drawing by King and Pantling (1898: pl. 6A) are a clear indication of intraspecific variability. The alternate explanation that the type gathering from Tropical Valley at CAL and P is composed of multiple, barely distinguishable species is too far-fetched.

Blatter and McCann (1931) compared O. *lingmalensis* only to O. *verticillata* from which it was distinguished by the flatter scape, the slightly erose linear bracts of variable length, long pedicelled ovary, and erose petals. The species has been considered a variety of O. recurva by Santapau and Kapadia (1960, 1966), Sharma et al. (1984) and Naithani (1990). The characters given by Blatter and McCann (1931) are congruent with O. brachystachys, specifically the vegetative description (no indication on jointed or unjointed leaves), the size of the flowers (2 mm), the shape of the bract, the erose petals, and the lip with erose lateral and epichile lobes. The species has never been illustrated. The holotype could not be examined and the repositories of the paratypes are unknown.

For *O. brachyphylla*, Blatter and McCann (1931) did not designate any type material. Their illustration was prepared from a live specimen, and the fate of that plant is unknown. The neotype designation by Santapau and Kapadia (1960) is in violation of Art. 9.19(a) of Shenzhen Code (Turland *et al.*, 2018), because the illustration in the protologue constitutes original material (Art. 9.4b). Any original material must be considered for lectotypification before a neotype can be designated. Accordingly, the neotype designation by Santapau and Kapadia (1960) is here superseded by the lectotypification of *O. brachyphylla* through the illustration of the protologue (Blatter & McCann, 1931: pl. 2).

Oberonia brachyphylla (Fig. 1k,l) and *O. brachystachys* share the small vegetative size, the lack of jointed leaves, the small flowers (1.5 mm), the oval petals with erose margins, and the four-lobed lip with erose margins of the lateral lobes, entire constricted mesochile, and erose edges of the epichile lobes. The plant illustrated by Blatter and McCann (1931) is somewhat smaller than the types of the other names. Those size and slight shape differences of the leaves fall well within the known variability in other species, particularly *O. rufilabris* Lindl. (Geiger, pers. obs.). Blatter and McCann (1931) compared their species only to *O. recurva* and neglected *O. brachystachys*. They distinguished *O.*

brachyphylla from *O. recurva* by the leaf shape (demonstrably highly variable), longer inflorescence (demonstrably highly variable in many species: Geiger, 2019), oblong sub-acute petals (no difference), lip much longer than the sepals (no difference), mid lobe square, bifid and incurved (no difference). Accordingly, the differentiation was based on lack of knowledge on intraspecific variability and outright errors, hence, *O. brachyphylla* is treated here as the synonym of *O. brachystachys*.

The alleged occurrence of *O. brachystachys* in China (Seidenfaden, 1968) is questionable (Chen *et al.*, 2009: as *O. recurva*). Given the past confusion of *O. brachystachys* and *O. subligaculifera* and their synonyms, Seidenfaden's (1968) indication may be based on *O. subligaculifera*, which would match distributional patterns of the two species better.

Mitra (1958) listed *Malaxis cordifolia* Rchb.f. as the correct name for *O. demissa*. There are two *M. cordifolia* listed in WCSP (2020): Sm. in A.Rees, 1812, and the *nom. illeg*. by (Rolfe) Ames & C. Schweinf. in O. Ames, 1920. The former is considered a synonym of *Liparis petiolata* (D.Don) P.F.Hunt & Summerh. It is not clear what name Mitra (1958) was referring to. In any case, any potential Reichenbach name cannot threaten *O. brachystachys*, as Reichenbach's orchid species names are all from after 1845 and placement in *Oberonia* would be after that.

AMES 72799/HUH 00101944 is listed in AMES on-line catalogue as a possible isotype of *O. brachystachys.* However, given the specific collector and gathering numbers cited by Lindley (1838), which are not present on the AMES sheet, that specimen has no type standing at all.

The variability of *O. brachystachys* seems extensive, some may even suspect excess lumping of taxa. However, based on available material and information, it is impossible to form distinct groupings that would segregate the plants consistently with more than one character. For instance, the degree of serration of the petals is not correlated with the degree of serration of the lip, and neither is correlated with shape of the epichile lobes on the lip. The variability of the sac from a small depression (Figs. 3k,n, 4b,e,f) to distinct cavity (Fig. 3c,d) is surprising. All other flower attributes of Figure 4 with minute sac and the live plant of Figure 3c,d with distinct sac are identical. Ansari and Balakrishnan (1990: fig. 32g, 35g, 36g-i) seem to have noticed different extents of the sac formation as indicated by the shading in the central upper portion of the lip. The combination of absence of other confirming characters and evidence of variability by Ansari and Balakrishnan (1990) suggest as the simplest explanation that the sac condition is variable in O. brachystachys. To demonstrate distinct groupings, much more material would need to be collected, and the morphological data would ideally be supported with molecular techniques. The latter approach has demonstrated extensive intraspecific variability in Oberonia species (Geiger et al., 2020).

Oberonia subligaculifera J.J.Sm., Bull. Jard. Bot. Buitenzorg, sér. 2, 9: 35. 1913. *Holotype*: INDONESIA, **Java**, Jawa Barat, near Tirtasari, 1500 m, *J.J. Smith & Rant* 201 (repository unknown).

Oberonia bambusicola Kerr, Bull. Misc. Inform. Kew 1927: 213. 1927. *Syntype:* THAILAND, Chon Buri, Sriracha [= Si Racha], Nang Khaw, 50 m, *Kerr* 383 (K [K000596096]!).

Oberonia evrardii Gagnep., Bull. Soc. Bot. France 76: 326. 1929. *Lectotype*: (inadvertent designation by Seidenfaden, 1968: 48 by reference to "P!, type"): VIETNAM, Lam ĐÑng, Dalat, chalet Rimaud, 28.10.1920, *Evrard* 300 (P [P00404954]!).

Oberonia kanburiensis Seidenf., Bot. Tidsskr. 68: 47. 1973. *Type*: THAILAND, **Kanchanaburi**, West of Siswat, Hauy Ban Kao, *Beusekom et al. 3657* (holo C (C2103) *n.v.*, iso L [L0061749]!).

Oberonia arunachalensis A.N.Rao, Rheedea 7: 130. 1997. *Type*: INDIA, **Arunachal Pradesh**, Lohit district, Kamlang Reserve forest on way to Chamba-glowlake area, 500 m, *A.N. Rao* 28067 (holo OHT *n.v.*). syn. nov.

Oberonia raoi L.R.Shakya & R.P.Chaudhary, Rheedea 10: 57. 2000 ["*raoii*"]. *Type*: INDIA, **Meghalaya**, Khasia hills, Nongpoh, 4000 ft. (1219 m), August 1897, *Pantling s.n.* (holo CAL [CAL0000024756]!). Erroneous citation of type gathering in protologue, see below. **syn. nov.**

Oberonia kamlangensis A.N.Rao, J. Econ. Taxon. Bot. 24: 267. 2000. Type: INDIA, Arunachal Pradesh, Lohit district, Kamlang sanctuary, 150 m, A.N. Rao 28283A (holo OHT *n.v.*); *ibid*, A.N. Rao 28283-B (para CAL *n.v.*); *ibid*. A.N. Rao 28283-C (para OHT *n.v.*). syn. nov. Figs. 687

Epiphytic herbs, usually 4-6 leaves, jointed, forming stemless fan. Leaves narrow elongated acuminate, acute, typically slightly falcate towards or away from growth axis, typically 2–8 cm long, 0.3–0.5 cm wide. Roots thin, fibrous, unbranched. Inflorescence typically 5-10 cm long, typically 2-3 × as long as leaves. Scape short, rachis longitudinally grooved, flowers variable arranged from distinct whorls, to in spiral, or scattered. Flowers 2.0–2.5 × 2.2–3.0 mm, average ~2.5 × 2.5 mm, green, yellow-orange, orange. Bracts, elongated acuminate, acute, erose, about as long as flower. Pedicelled ovary round, ~2.0–2.5 mm long. Orientation of all tepals somewhat forward or somewhat recurved. Sepals broad oval acute. Petals oblong obtuse, erose to dentate or laciniate. Lip four-lobed, lateral lobes square or trapezoidal, margin erose to dentate; mesochile more or less constricted, entire; epichile lobes truncated, rounded or sub-quadrate, margins dentate or erose, disc indistinct, sac unknown.

Flowering: Flowering throughout year with peaks in February and October (Fig. 7a). The gap in June– July is considered a data artifact.

Habitat: Erect or pendulous epiphyte, evergreen to deciduous, broad-leaf, mixed and conifer forest, on branches of tree and shrubs, on bamboo, in coffee plantation, 0–2400 m, predominantly below 900 m (Fig. 7a).

Distribution: India, Malaysia, Myanmar, Thailand, Vietnam, China, Indonesia, Philippines (Luzon) (Kerr, 1927; Santapau & Kapadia, 1960; Backer & Bakhuizen van der Brink, 1968; Seidenfaden, 1968, 1978; Saldanha in Saldanha & Nicolson, 1976; Pradhan, 1979; Abraham & Vatsala, 1981; Manilal, 1988; Comber, 1990; Naithani, 1990; Sakkar, 1995a; Turner, 1995; Hop, 1998; Bose *et al.*, 1999; Averyanov & Averyanova, 2003; Kumar & Manilal, 2004; Rao, 2010; Averyanov, 2013; Bunpha *et al.*, 2013, 2019; Jin *et al.*, 2019; herbarium records).

Specimens examined: INDONESIA, Java, Bandong, Zollinger 897 (P[P00428170: left]). Sumatra, Gunung Leuser Nature Reserve, Atjeh, Ketambe valley of Lau Alas, near tributary of Lau Ketambe, de Wilde & de Wilde-Duyfjes 12267 (K). MALAYSIA, Sarawak, Marudidistrict between Bario and Pa Umor, Beaman 11256 (K). MYANMAR, King Lung, MacGregor 822 (E [E00616243]). PHILIPPINES, Luzon, Cordillera Administrative Region, Bontoc. Vanoverbergh 3828 (F [F452930], MO [MO799960]). THAILAND, Mae Hong Son, Khun Yuam, Larsen & Larsen 34115 (K). VIETNAM, Ha Lang, Thanh Nhat, 42 km E of Cao Bang, Averyanov CLB 732 (MO [MO04970868]). Noh Quan, Cuc Phuong National Park, CP146; Cuong et al. 775 (MO [MO5336640]).

Notes: The repository of the holotype of *O. subligaculifera* J.J.Sm. is unknown. The specimen is a holotype as the protologue specified that it was based on a single specimen. The species (Fig. 6a) and its previous recognized synonyms (Geiger, 2019) *O. evrardii* (Fig. 6d,e) and *O. kanburiensis* (Fig. 6 f, g) is characterized by the elongate acuminate jointed leaves, the larger flowers $(1.8-2.5 \times 2.2-$ 3.0 mm), petals with strongly erose to incised margins, the four lobed lip with strongly erose to incised lateral lobes, a constriction of the entire mesochile, and a bifid epichile with erose to incised tips. Although the flowers are very similar to those of *O. brachystachys*, the jointed leaves of *O.*

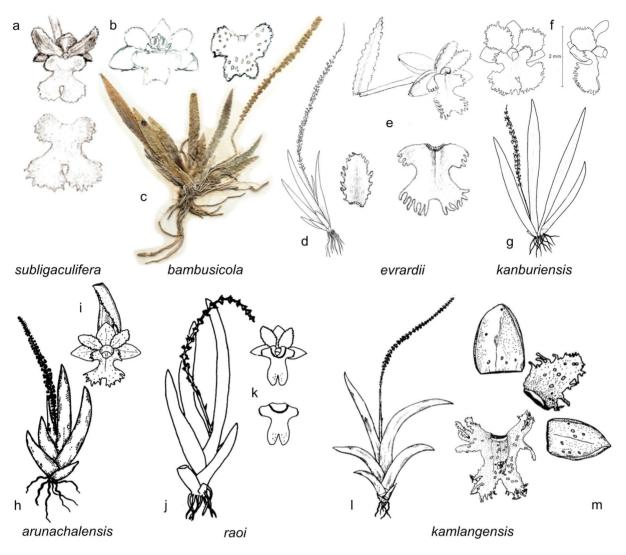


Fig. 6. Drawings and photographic image of *Oberonia subligaculifera* J.J.Sm. and its synonyms: **a**. *O. subligaculifera* illustration by J.J. Smith from Schuiteman and de Vogel (2006), entire flower and isolated lip; **b**,**c**. *O. bambusicola* Kerr 383 (syntype K [K000596096]); **b**. Drawing of flower and lip on note affixed to type sheet; **c**. Habit; **d**,**e**. *O. evrardii Gangnep.*; **d**. Drawing of lectotype *Evrard* 300 (P [P00404954]) by Seidenfaden (1968: fig. 27); **e**. Drawing on lectotype sheet of entire flower, isolated lip, and isolated petal; **f**,**g**. *O. kanburiensis* Seidenf. drawing of type from Seidenfaden (1978: fig. 9); **f**. Flower; **g**. Habit; **h**,**i**. *O. arunachalensis A.N. Rao* drawing from Rao (1997: fig. 1); **h**. Habit; **i**. Flower; **j**,**k**. *O. raoi* L.R. Shakya and R.P. Chaudhary drawing from Shakya and Chaudhary (2000: fig. 1); **j**. Habit; **k**. Flower; **l**,**m**. *O. kamlangensis* A.N. Rao drawings from Rao (2000: pl. 1); **l**. Habit; **m**. Flower parts digitally rearranged to natural positions; (a, d, f–m from cited publications under fair use provision of copyright law; photos b,c,e by D.L. Geiger).

subligaculifera (unjointed in *O. brachystachys*) in combination with the typically larger flowers indicate that they are distinct species. While intraspecific variability is extensive in *Oberonia*, the jointed vs. unjointed leaves have never been suggested as variable within species. On the contrary, that character has been used to separate the genus *Oberonia* into two subgenera (Schlechter, 1911): *Oberonia* s.s. [= *Apothemnophyllum* Schtr.] with jointed leaves, and *Menophyllum* Schltr. with unjointed leaves. Bunpha *et al.* (2019) considered *O. evrardii* distinct from *O. subligaculifera* based on differences in the depression of the lip and the shape of the rostellum. Neither of those characters were illustrated, and the sample size for those two species were four and five, respectively. The difference is at best subtle, with significant intraspecific variability demonstrated previously (Geiger, 2019). Those characters are considered insignificant. The shape of the rostellum is likely

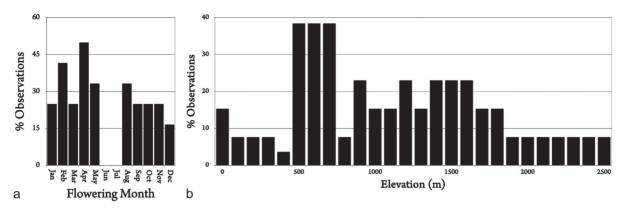


Fig. 7. Summary bar charts of ecological data for Oberonia subligaculifera J.J.Sm., compiled from any of its synonyms: **a.** Phenology data from Smith (1913), Santapau and Kapadia (1960), Saldanha and Nicolson (1976), Abraham and Vatsala (1981), Manilal (1988), Ansari & Balakrishnan (1990), Nayar *et al.* (2006), Bunpha *et al.* (2013, 2019), Jin *et al.* (2019), herbarium records; **b.** Elevation data from Smith (1913), Kerr (1927), Ansari and Balakrishnan (1990), Seidenfaden (1992), Rao (1997), Averyanov *et al.* (2000), Averyanov and Averyanova (2003), Averyanov (2013), Bunpha *et al.* (2013, 2019), Jin *et al.* (2019), herbarium records; (graphics by D.L. Geiger).

affected by ontogeny and state of maturation, hence, is meaningless.

Oberonia bambusicola Kerr (Fig. 6b,c) had been considered a synonym of *O. brachystachys* (Seidenfaden, 1968; WCSP, 2020), but is here placed in synonymy with *O. subligaculifera*. The shared characters between *O. bambusicola* and *O. subligaculifera* include the jointed leaves (visible in K syntype), the size of the flowers (>3 mm), the greenish flower color (in fluid preserved specimen), the erose petals, and all aspects of the lip. It differs from *O. brachystachys* by the jointed vs. unjointed leaves, the larger flowers (>3 mm vs. ~1.5 mm), and the flower was not given by Kerr (1927), it can be inferred by adding his measurements of the lip (2.2 mm long) and the sepal (1.5 mm long).

Kerr (1927) compared his species only to *O. recurva* and noted the proportionally shorter lip and the less erose petals. The annotations by Kerr preserved on the type sheet identify his specimen as cf. *O. brachystachys*, a further indication to the synonymy of *O. brachystachys* and *O. recurva* discussed above. He did not mention *O. subligaculifera*.

Oberonia arunachalensis (Fig. 6h,i) is a synonym of *O. subligaculifera*. It shares the fan-shaped habit, the jointed leaves (according to protologue, not evident in figure), the oval and erose petals, and the lip shape

with serrated lateral lobes and epichile lobes with *O. subligaculifera*. The two names refer to the same species. Rao (1997) compared his *O. arunachalensis* only with *O. kanburiensis*, and distinguished the two by the longer leaves and inflorescence (both extensively variable within species), green flowers (no difference), elliptical-elongate petals (no difference) and dentate lobes of the lip (no difference). Accordingly, the differentiation was based on lack of knowledge of intraspecific variability and outright mistakes.

The flowers appear to be rather small (c. 1.7×1.4 mm) based on the scale bar associated with the flower. Errors in scale bars and drawings are abundant in micro-floral orchids (Geiger, 2019). According to the scale bar of the entire flower (Rao, 1997: fig. 1d) the bract is 1.3 mm long, while the isolated bract shown in Rao (1997: fig. 1e) is 1.5 mm long based on its scale bar. Whether this should illustrate variability of floral parts on the same inflorescence or whether those are scale bar errors is unclear. Even with a 15% variance in measurement accuracy, the flowers of O. arunachalensis would be rather small for O. subligaculifera and more compatible with O. brachystachys. Intraspecific variability of floral size by up to 50% have been documented in other species (O. complanata: Geiger, 2019; O. equitans: Geiger et al., 2020); O. arunachalensis falls within

that documented range of intraspecific variability with respect to *O. subligaculifera*. The jointed leaves according to protologue, not shown in Rao (1997: figs. 1c; fig. 6h), in addition to the more eastern type locality is indicative of a synonymy with *O. subligaculifera*.

The protologue of *O. raoi* misspelled the specific epithet as *raoii*, an error already noted by Badyopdhyay *et al.* (2016). The protologue further mistook an old herbarium ledger number [449798] as the gathering number of the type; the correct citation of the gathering is *Pantling s.n.*

Oberonia raoi (Fig. 6j,k) is a synonym of O. subligaculifera. The common characters include the habit with jointed leaves, the erose margins of the ovate petals, the overall shape of the lip, and the erose margin of the square lateral lobes at right angle to the axis of the lip. The only character not matching are the margins of the apical lobes of the lip, which are distinctly erose in O. brachystachys while appearing entire in the drawing of O. raoi. However, the quality of the drawing is not particularly good and one may wonder whether artistic license missed those small details. Given the large number of matching details including the jointed leaves, O. raoi is here synonymized under O. subligaculifera. Oberonia raoi was only compared to O. myriantha (= O. acaulis) and not to O. subligaculifera.

Oberonia kamlangensis (Fig. 6l,m) is a synonym of *O. subligaculifera*. It was compared to *O. mucronata* (D.Don) Ormerod & Seidenf. in G.Seidenf., but not to *O. subligaculifera* or to *O. arunachalensis* described by the same author three years earlier (Rao, 2000). It is another case where a new species is justified by comparison to a very distant and different species while the more similar species were not considered. *Oberonia kamlangensis* is indistinguishable from *O. arunachalensis*, itself a synonym of *O. subligaculifera*. The jointed leaves, the strongly erose petals, and the lip with strongly laciniate lateral lobes and the equally deeply incised epichile lobes (Fig. 6h,l,m) are shared by the two nominal species.

The flower color was given as pale yellow-green by Smith (1913), but the description was based on ethanol-preserved material. Ethanol bleaches the flower color (Geiger, pers. obs.) with orange flowers turn yellow green. Green flowers have been reported by Rao (1997: as *O. arunachalensis*). Variability in flower color from green to orange is well-known in *O. mucronata* (Geiger, 2019).

Discussion

The untangling of the two species discussed here leads to a consistent overall pattern. The names are distinguished by size of the flowers (1.0–1.75 mm in *O. brachystachys*, 1.8–3.0 mm in *O. subligaculifera*), the un-jointed leaves in *O. brachystachys*, but jointed ones in *O. subligaculifera*. *Oberonia brachystachys* is distributed from western and southern India, including Sri Lanka to Myanmar, while *O. subligaculifera* is found from easternmost India to Java and the Philippines.

The synonymy of O. parvula/croftiana under O. brachystachys may appear too sweeping. The limited sample size, the documented intermediate forms, and the known extensive intraspecific variability demonstrated with molecular techniques (Li et al., 2016; Geiger et al., 2020) support our conclusion as the best explanation of all data. It would be worth-while to investigate the matter further with combined floral (z-stack light microscopy, SEM) and molecular data of the same specimens exhibiting a range of morphologies. It might be the case that the *parvula*-morph of O. *brachystachys* is a species in statu nascendi, i.e., that those specimens represent lineages that are in the process of diverging. However, the divergence does not seem to be complete, and does not need to be recognized taxonomically at this point in time.

As shown in other cases, the microfloral *Oberonia* were over-described due to lack of knowledge of intraspecific variability, focussing on minute differences of individual plants rather than applying population thinking in systematics, citing characters as differences that are in fact identical, and comparing new names to highly

dissimilar species rather than the more similar ones. The last factor may also be caused by lack of knowledge of obscure plant groups. Ideally, in order to describe a new species in a genus, every name ever described in this genus has to be understood by the proposer of the new name. That entails a revision of the entire genus, either already published, or carried out by the describer. Obscure groups by their very nature have not been revised. Accordingly, describing new species in obscure plant group has a high potential of introducing synonyms (Geiger, 2021).

Geiger *et al.* (2020) additionally discussed the higher academic recognition for introducing new names as opposed to recognizing synonyms, and the lack of a malus for burdening systematics with excess names. We here discuss additional examples showing multiple incongruent drawings from the same specimen, while providing superior photographic images and scanning electron micrographs.

Focus on political geographic units is a further contributing factor. A case in point is the Indian synonyms of *O. subligaculifera* described from Indonesia. The dispersal potential of *Oberonia* spp. is extensive due to their minute seeds (Bartholott *et al.*, 2014; Geiger, 2014, 2020, unpubl. data; Geiger *et al.*, 2020), and has been confirmed by molecular data (Geiger *et al.*, 2020). Accordingly, new distributional records of already described species are too often mistaken as new species. Postulated microendemic *Oberonia* species (Averyanov, 2013; Averyanov *et al.*, 2015, 2019) are most likely synonyms of already described species (Geiger, 2020, 2021, unpubl. data).

Acknowledgements

The authors are grateful to Ministry of Environment, Forest & Climate Change for funding the Flora of India Project; Dr. A.A. Mao, Director, Botanical Survey of India, Kolkata, Dr. M.U. Sharief, Head of Office, Botanical Survey of India, Southern Regional Centre, Coimbatore for the providing the facilities and support to carry out the work; Dr. K. Karthigeyan (CNH) for consulting herbarium and to Kerala Forest Department for granting the field permission. DLG was accommodated during various collection visits at the following herbaria: Jacek Wajer (BM), Hanna Atkins and Suzanne Cubey (E), Christine Niezgoda (F), André Schuiteman (K), Wayne Gerbet (MEL), James Solomon and Donna Herrera (MO), Marc Pignal (P), Rusty Russell (US), Ernst Vitek (W), Walter Till (WU). The reviewers and editor helped to improve the manuscript.

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