

Supplementary Material S1: References of Progress on Flora treatments for the family Convolvulaceae, organized by region or country.

- ACEVEDO-RODRÍGUEZ P. & M.T. STRONG 2012. Catalogue of seed plants of the West Indies. *Smithsonian Contributions to Botany* 98: 1–1192.
- ACEVEDO-RODRÍGUEZ P. 2005. *Vines and Climbing Plants of Puerto Rico and the Virgin Islands*. Department of Botany, National Museum of Natural History.
- ALLAN HERBARIUM 2023. *Checklist of the New Zealand flora - seed plants*. Plant Names Database. <https://biotanz.landcareresearch.co.nz/checklists#:~:text=Seed%20plants>
- AUSTIN D.F. 1990. Convolvulaceae. In: WAGNER, W.L., HERBST, D.R. & S.H. SOHMER (eds.), *Manual of the Flowering Plants of Hawaii*. Volume 1. University of Hawaii Press & Bishop Museum Press, Honolulu. pp. 550–564.
- BAASANMUNKH S., URGAMAL M., OYUNTSETSEG B., SUKHORUKOV A.P., TSEGMEZ Z., SON D.C., ERST A., OYUNDELGER K., KECHAYKIN A.A., NORRIS J., KOSACHEV P., MA J.S., CHANG K.S. & H.J. CHOI 2022. Flora of Mongolia: Annotated checklist of native vascular plants. *PhytoKey* 192: 63– 169. <https://doi.org/10.3897/phytokeys.192.79702>
- CARRANZA E. 2007. Familia Convolvulaceae. In: RZEDOWSKI J. & G. CALDERÓN DE RZEDOWSKI (eds.), *Flora del Bajío y de Regiones Adyacentes*. Fascículo 151. Instituto de Ecología, A.C., Centro Regional del Bajío, Pátzcuaro, Michoacán, México. pp. 1–129.
- CARRANZA E. 2008. *Convolvulaceae II*. In: RZEDOWSKI J. & G. CALDERÓN DE RZEDOWSKI (eds.), *Flora del Bajío y de Regiones Adyacentes*. Volume 2, Fascículo 155, Instituto de Ecología, A.C., Centro Regional del Bajío, Pátzcuaro, Michoacán, México. pp. 1–107.
- COSTEA M., FELGER R.S., AUSTIN D.F., VAN DEVENDER T.R. & J.J. SÁNCHEZ-ESCALANTE 2012. *Convolvulaceae* of Sonora, Mexico. II. *Cuscuta*. *Journal of the Botanical Research Institute of Texas* 6(2): 527–548.
- D'ARCY W.G. 1987. *Flora of Panama: Checklist and Index*. Missouri Botanical Garden, St. Louis. 358 pp.
- DAVIS B., COUCH C., BIDAULT E., SIMBIANO F.J. & D. MOUMOU 2024. *Convolvulaceae of Guinea: Taxonomy, conservation, and useful plants*. *BioRxiv* 2024.07.15.602708 <https://doi.org/10.1101/2024.07.15.602708>
- FELGER R.S., VAN DEVENDER T.R., AUSTIN D.F., SÁNCHEZ-ESCALANTE J.J. & M. COSTEA 2012. *Convolvulaceae* of Sonora, Mexico. I. *Convolvulus*, *Cressa*, *Dichondra*, *Evolvulus*, *Ipomoea*, *Jacquemontia*, *Merremia*, and *Operculina*. *Journal of the Botanical Research Institute of Texas*, 6(2): 459–527.
- FOSBERG F.R. & M.H. SACHET 1977. Flora of Micronesia, 3: Convolvulaceae. *Smithsonian Contributions to Botany* 36: 1–38. <https://doi.org/10.5479/si.0081024X.36>
- GONÇALVES M.L. 1987. Convolvulaceae. In: LAUNERT E. (ed.), *Flora Zambesiaca*. Flora Zambesiaca Managing Committee.
- GREUTER W., RANKIN RODRÍGUEZ R. & RAAB-STRAUPE, E. VON (eds.) 2017. *Flora de la República de Cuba*. Botanischer Garten und Botanisches Museum Berlin-Dahlem & Jardín Botánico Nacional de Cuba.
- HAMMEL B.E. 2010. Convolvulaceae. In HAMMEL B.E., GRAYUM M.H., HERRERA C. & N. ZAMORA (eds.) *Manual de Plantas de Costa Rica, Volume V: Dicotiledóneas (Clusiaceae–Gunneraceae)*. *Monographs in Systematic Botany from the Missouri Botanical Garden*, 119: 72–126.
- HUTCHINSON J. & J.M. DALZIEL 1952. *Flora of West Tropical Africa*. Crown Agents for Oversea Governments and Administrations.
- JØRGENSEN P.M. & S. LEÓN-YÁNEZ (eds.). 1999. *Catalogue of the vascular plants of Ecuador*. Missouri Botanical Garden Press, Missouri.
- KOMAROV V.L. (ed.). 1969. *Flora of the U.S.S.R.* Israel Program for Scientific Translations, Springfield.
- LIPSCOMB B. 2023. *Flora of North America North of Mexico*. Volume 14, Magnoliophyta: Gentianaceae to Hydroleaceae. *Journal of the Botanical Research Institute of Texas* 17(2): 540–540.
- MAQUET P. 1985. Convolvulaceae. In: TROUPIN G. (ed.), *Flore du Rwanda: Spermatophytes*, Volume 3. Musée royal de l'Afrique centrale, Rwanda. pp. 231–254.
- MCDONALD A. 1993. *Flora de Veracruz*. Fascículo 73: Convolvulaceae I. Instituto de Ecología/University of California, Veracruz. pp.1–99. <https://doi.org/10.21829/fv.408.1993.73>
- MCDONALD A. 1994. *Flora de Veracruz*. Fascículo 73: Convolvulaceae II. Instituto de Ecología/University

- of California, Veracruz. pp. 1–133. <https://doi.org/10.21829/fv.404.1994.77>
- MCPHERSON G. & D.F. AUSTIN 1993. Convolvulaceae. In: BRAKO L. & J.L. ZARUCCHI (eds.), *Catalogue of the Flowering Plants and Gymnosperms of Peru*, pp. 365–374. Missouri Botanical Garden, St. Louis.
- MEEUSE A.D.J. & W.G. WELMAN 2000. *Flora of Southern Africa: Convolvulaceae*, Volume 28(1). National Botanical Institute, Pretoria.
- MUNZINGER J. & T. DEROIN 2001. *Diegodendraceae, Convolvulaceae*. In: HUMBERT H. & J.-F. LEROY (eds.), *Flore de Madagascar et des Comores: plantes vasculaires*. Muséum National d'Histoire Naturelle, Paris, 287 pp.
- MWANGA MWANGA I.J.C., SOSEF M.S.M. & A.R.G. SIMÕES 2022. *Flore d'Afrique Centrale: Spermatophytes, Convolvulaceae*. National Botanic Garden of Belgium, 254 pp.
- O'DONELL C.A. 1959. Convolvuláceas argentinas. *Lilloa* 29: 87–348.
- O'DONELL C.A. 1959. Convolvuloideas de Uruguay. *Lilloa* 29: 349–376.
- OOSTSTROOM S.J. VAN. 1953. Convolvulaceae. In: STEENIS C.G.G.J. VAN (ed.). *Flora Malesiana*. Series 1, Volume 4, Noordhoff-Kolff, Djakarta. pp. 388–512.
- PALACIOS-WASSENAAR O. & G. CASTILLO-CAMPOS 2024. *Flora de Veracruz*. Fascículo 190: *Convolvulaceae III. Cuscuta*. Instituto de Ecología. University of California, Xalapa, Veracruz. <https://doi.org/10.21829/fv.551.2021.190>
- PELSER P.B., BARCELONA J.F. & D.L. NICKRENT (eds.). 2011. *Co's Digital Flora of the Philippines*. www.philippineplants.org
- RHUI-SHENG F. & G. STAPLES 1995. Convolvulaceae. In: WU, Z.Y. & P.H. RAVEN (eds.), *Flora of China*, Volume 16 (Gentianaceae through Boraginaceae). Science Press, Beijing, and Missouri Botanical Garden Press, St. Louis. pp. 271–325.
- RODRÍGUEZ R., MARTICORENA C., ALARCÓN D., BAEZA C., CAVIERES L., FINOT V.L., FUENTES N., KIESSLING A., MIHOC M. & A. PAUCHARD 2018. *Catálogo de las plantas vasculares de Chile*. *Gayana Botánica* 75(1): 1–430. <https://doi.org/10.4067/S0717-66432018000100001>
- RZEDOWSKI J. & E. CARRANZA GONZÁLEZ 2023. Sinópsis de la familia Convolvulaceae en México (Synopsis of the family Convolvulaceae in Mexico). *Journal of the Botanical Research Institute of Texas* 17(1): 271–279. <https://doi.org/10.17348/jbrit.v17.i1.1296>
- RZEDOWSKI J. 2005. *Flora Fanerogámica del Valle de México*. Instituto de Ecología, Veracruz.
- SANTAPAU H. & A.N. HENRY 1973. *A Dictionary of the Flowering Plants in India*. CSIR, New Delhi.
- SIMÃO-BIANCHINI R., FERREIRA P.P.A., PASTORE M., DELGADO-JUNIOR G.C., VASCONCELOS L.V., PETRONGARI F.S., MOREIRA A.L.C., BURIL M.T., SIMÕES A.R.G. & C.V. SILVA 2020. *Flora do Brasil 2020*. Jardim Botânico do Rio de Janeiro. <https://floradobrasil2020.jbrj.gov.br/FB93>
- SOSEF M.S.M., FLORENCE J., NGOK BANAK L. & H.P. BOUROBOU (eds.) 2015. *Flore du Gabon, Volume 46: Convolvulaceae, Passifloraceae*. Margraf Publishers, Weikersheim. <https://doi.org/10.5281/zenodo.1107274>
- STANDLEY P.C. & L.O. WILLIAMS 1970. Convolvulaceae. In: STANDLEY P.C. & L.O. WILLIAMS (eds.), *Flora of Guatemala - Part IX. Fieldiana: Botany* 24(9): 4–85.
- STAPLES G. & S. SYAHIDA-EMIZA 2015. Convolvulaceae. In: KIEW R., CHUNG R.C.K., SAW L.G. & E. SOEPADMO (Eds.), *Flora of Peninsular Malaysia, Series II: Seed Plants*, Volume 5. Forest Research Institute Malaysia (FRIM), pp. 55–198.
- STAPLES G. 2010. Convolvulaceae. In: SANTISUK T. & K. LARSEN (Eds.). *Flora of Thailand*, 10: 330–468.
- STAPLES G. 2018. *Flora of Cambodia, Laos and Vietnam (formerly Flore du Cambodge, Laos et du Vietnam)*, Volume 36: Convolvulaceae. National Museum of Natural History, Paris. 407 pp.
- STAPLES G.W. & S.Z. YANG 1998. Convolvulaceae. In: HUANG T.C., et al. (eds.), *Flora of Taiwan*, 2nd Edition. Department of Botany, National Taiwan University, Taipei, pp. 341–384.
- THULIN M. (ed.), *Flora of Somalia*. Volume 3. The Royal Botanic Gardens, Kew. pp. 221–258.
- VERDCOURT B. 1963. *Flora of Tropical East Africa: Convolvulaceae*. Crown Agents for Overseas Governments & Administrations, London. 161 pp.
- WOOD J.R.I., BIANCHINI R. & A. FUENTES 2015. Convolvulaceae. In: JØRGENSEN P.M., NEE M.H. & S.G. BECK (eds.), *Catálogo de las plantas vasculares de Bolivia*. pp. 520–531. Missouri Botanical Garden Press.

Supplementary Table S2: List Convolvulaceae species with Genomic accessions in Genbank

Species	Type of Genome	Accession Number (s)	Citation
<i>Argyreia nervosa</i> (Burm.f.) Bojer	Chloroplast	KF242477	Eserman <i>et al.</i> (2014)
<i>Argyreia nervosa</i> (Burm.f.) Bojer	Chloroplast	KF242477	Eserman <i>et al.</i> (2014)
<i>Argyreia strigillosa</i> C.Y.Wu	Chloroplast	OR770998	Chen <i>et al.</i> (2024)
<i>Argyreia velutina</i> C.Y.Wu	Mitochondrial	MZ240724	Lin <i>et al.</i> (2022)
<i>Calystegia soldanella</i> (L.) Roem. & Schult.	Mitochondrial	MZ240725	Lin <i>et al.</i> (2022)
<i>Convolvulus arvensis</i> L.	Mitochondrial	BK059236	Lin <i>et al.</i> (2022)
<i>Convolvulus tragacanthoides</i> Turcz.	Chloroplast	OR778088	Chen <i>et al.</i> (2024)
<i>Cuscuta africana</i> Willd.	Chloroplast	MN464179	Banerjee and Stefanović (2020)
<i>Cuscuta australis</i> R.Br.	Chloroplast	OR778086	Chen <i>et al.</i> (2024)
<i>Cuscuta australis</i> R.Br.	Mitochondrial	BK059204	Anderson <i>et al.</i> (2021)
<i>Cuscuta australis</i> R.Br.	Nuclear	PRJNA394036	Sun <i>et al.</i> (2018)
<i>Cuscuta approximata</i> Bab.	Chloroplast	MN464180	Banerjee and Stefanović (2020)
<i>Cuscuta bonafortunae</i> Costea & I.García	Chloroplast	MK887215	Banerjee and Stefanović (2019)
<i>Cuscuta boldinghii</i> Urb.	Chloroplast	MK881074	Banerjee and Stefanović (2019)
<i>Cuscuta campestris</i> Yunck.	Chloroplast	ENA: PRJEB19879	Chen <i>et al.</i> (2024)
<i>Cuscuta campestris</i> Yunck.	Mitochondrial	BK016277	Anderson <i>et al.</i> (2021)
<i>Cuscuta carnosa</i> Costea & I.García	Chloroplast	MK887212	Banerjee and Stefanović (2019)
<i>Cuscuta chapalana</i> Yunck.	Chloroplast	MK887214	Banerjee and Stefanović (2019)
<i>Cuscuta chinensis</i> Lam.	Chloroplast	MH780079, OL752638, OR771000	Park <i>et al.</i> (2019), Pan <i>et al.</i> (2023), Chen <i>et al.</i> (2024)
<i>Cuscuta costaricensis</i> Yunck.	Chloroplast	MK881072	Banerjee and Stefanović (2019)
<i>Cuscuta epilinum</i> Weihe	Mitochondrial	BK059237	Lin <i>et al.</i> (2022)
<i>Cuscuta epithimum</i> (L.) L.	Chloroplast	OP620588	Pan <i>et al.</i> (2023)
<i>Cuscuta erosa</i> Yunck.	Chloroplast	MK881073	Banerjee and Stefanović (2019)
<i>Cuscuta europaea</i> L.	Chloroplast	OP620589, OR770999, OR771001	Pan <i>et al.</i> (2023), Chen <i>et al.</i> (2024)
<i>Cuscuta europaea</i> L.	Mitochondrial	BK059238	Lin <i>et al.</i> (2022)
<i>Cuscuta exaltata</i> Engelm.	Chloroplast	EU189132	McNeal <i>et al.</i> (2007)

Species	Type of Genome	Accession Number (s)	Citation
<i>Cuscuta gronovii</i> Willd. ex Schult.	Chloroplast	AM711639, OL752639	Funk <i>et al.</i> (2007), Pan <i>et al.</i> (2023)
<i>Cuscuta japonica</i> Choisy	Chloroplast	OL752640, OR771002 - OR771012, MH780080	Park and Moon (2019), Pan <i>et al.</i> (2023), Chen <i>et al.</i> (2024)
<i>Cuscuta japonica</i> Choisy	Mitochondrial	MZ240726	Lin <i>et al.</i> (2022)
<i>Cuscuta mexicana</i> Yunck.	Chloroplast	MK887213	Banerjee and Stefanović (2019)
<i>Cuscuta nitida</i> E.Mey. ex Choisy	Chloroplast	MN464178	Banerjee and Stefanović (2020)
<i>Cuscuta obtusiflora</i> Kunth	Chloroplast	EU189133	McNeal <i>et al.</i> (2007)
<i>Cuscuta pedicellata</i> Ledeb.	Chloroplast	MN464181	Banerjee and Stefanović (2020)
<i>Cuscuta pentagona</i> Engelm.	Chloroplast	MH121054, OR771013, OR771014	Park <i>et al.</i> (2018), Chen <i>et al.</i> (2024)
<i>Cuscuta reflexa</i> Roxb.	Chloroplast	OR771015, OR771016	Chen <i>et al.</i> (2024)
<i>Cuscuta strobilacea</i> Liebm.	Chloroplast	MK867795	Banerjee and Stefanović (2019)
<i>Dichondra micrantha</i> Urb.	Chloroplast	OR771017	Chen <i>et al.</i> (2024)
<i>Dinetus duclouxii</i> (Gagnep. & Courchet) Staples	Chloroplast	OR771018	Chen <i>et al.</i> (2024)
<i>Dinetus dinetoides</i> (C.K.Schneid.) Staples	Chloroplast	OR771021	Chen <i>et al.</i> (2024)
<i>Dinetus racemosus</i> (Roxb.) Sweet	Mitochondrial	MZ240727	Lin <i>et al.</i> (2022)
<i>Distimake quinquefolius</i> (L.) A.R.Simões & Staples	Chloroplast	KF242501	Eserman <i>et al.</i> (2014)
<i>Erycibe obtusifolia</i> Benth.	Mitochondrial	MZ240728	Lin <i>et al.</i> (2022)
<i>Evolvulus alsinoides</i> (L.) L.	Mitochondrial	NC058741	Lin <i>et al.</i> (2022)
<i>Evolvulus nummularius</i> (L.) L.	Chloroplast	OR771019	Chen <i>et al.</i> (2024)
<i>Ipomoea alba</i> L.	Chloroplast	ON209203	Sudmoon <i>et al.</i> (2024)
<i>Ipomoea amnicola</i> Morong	Chloroplast	KF242478	Eserman <i>et al.</i> (2014)
<i>Ipomoea aquatica</i> Forssk.	Mitochondrial	MZ240730	Lin <i>et al.</i> (2022)
<i>Ipomoea argillicola</i> R.W.Johnson	Chloroplast	KF242479	Eserman <i>et al.</i> (2014)

Species	Type of Genome	Accession Number (s)	Citation
<i>Ipomoea batatas</i> (L.) Lam.	Nuclear	PRJEB14638, PRJNA301667	Yang <i>et al.</i> (2017)
<i>Ipomoea batatas</i> (L.) Lam.	Mitochondrial	OL699988	Yang <i>et al.</i> (2022)
<i>Ipomoea batatas</i> (L.) Lam.	Chloroplast	KF242473, KF242474, KF242475	Eserman <i>et al.</i> (2014)
<i>Ipomoea biflora</i> (L.) Pers.	Mitochondrial	MZ240723	Lin <i>et al.</i> (2022)
<i>Ipomoea cairica</i> (L.) Sweet	Chloroplast	KF242480	Eserman <i>et al.</i> (2014)
<i>Ipomoea cordatotriloba</i> Dennst.	Chloroplast	KF242497	Eserman <i>et al.</i> (2014)
<i>Ipomoea cordatotriloba</i> Dennst.	Chloroplast	MH173254	Chen <i>et al.</i> (2024)
<i>Ipomoea cynanchifolia</i> Meisn.	Chloroplast	MH173253	Chen <i>et al.</i> (2024)
<i>Ipomoea diamantinensis</i> J.M.Black ex Eardley	Chloroplast	KF242481	Eserman <i>et al.</i> (2014)
<i>Ipomoea dumetorum</i> Willd.	Chloroplast	KF242482	Eserman <i>et al.</i> (2014)
<i>Ipomoea eriocarpa</i> R.Br.	Chloroplast	KF242483	Eserman <i>et al.</i> (2014)
<i>Ipomoea hederifolia</i> L.	Chloroplast	KF242484	Eserman <i>et al.</i> (2014)
<i>Ipomoea involucrata</i> P.Beauv.	Chloroplast	KF242485	Eserman <i>et al.</i> (2014)
<i>Ipomoea lacunosa</i> L.	Chloroplast	MH173257	Chen <i>et al.</i> (2024)
<i>Ipomoea leucantha</i> Jacq.	Chloroplast	MH173263	Chen <i>et al.</i> (2024)
<i>Ipomoea minutiflora</i> (M.Martens & Galeotti) House	Chloroplast	KF242498	Eserman <i>et al.</i> (2014)
<i>Ipomoea murucoides</i> Roem. & Schult.	Chloroplast	KF242486	Eserman <i>et al.</i> (2014)
<i>Ipomoea nil</i> (L.) Roth	Chloroplast	KF242487	Eserman <i>et al.</i> (2014)
<i>Ipomoea nil</i> (L.) Roth	Chloroplast	AP017304	Hoshino <i>et al.</i> (2016)
<i>Ipomoea nil</i> (L.) Roth	Mitochondrial	NC031158	Yang <i>et al.</i> (2022)
<i>Ipomoea nil</i> (L.) Roth	Nuclear	BDFN01000001- BDFN01003416	Hoshino <i>et al.</i> (2016)
<i>Ipomoea obscura</i> (L.) Ker Gawl.	Chloroplast	KF242499	Eserman <i>et al.</i> (2014)
<i>Ipomoea obscura</i> (L.) Ker Gawl.	Chloroplast	OR995405	Sudmoon <i>et al.</i> (2024)
<i>Ipomoea orizabensis</i> (G.Pelletan) Ledeb. ex Steud.	Chloroplast	KF242488	Eserman <i>et al.</i> (2014)

Species	Type of Genome	Accession Number (s)	Citation
<i>Ipomoea pedicellaris</i> Benth.	Chloroplast	KF242489	Eserman <i>et al.</i> (2014)
<i>Ipomoea pes-caprae</i> (L.) Br.	Chloroplast	KF242490	Eserman <i>et al.</i> (2014)
<i>Ipomoea pes-tigridis</i> L.	Chloroplast	KF242500	Eserman <i>et al.</i> (2014)
<i>Ipomoea polpha</i> R.W. Johnson	Chloroplast	KF242491	Eserman <i>et al.</i> (2014)
<i>Ipomoea quamoclit</i> L.	Mitochondrial	MZ240732	Lin <i>et al.</i> (2022)
<i>Ipomoea setosa</i> Ker Gawl.	Chloroplast	KF242492	Eserman <i>et al.</i> (2014)
<i>Ipomoea splendor-sylvae</i> House	Chloroplast	KF242493	Eserman <i>et al.</i> (2014)
<i>Ipomoea splendor-sylvae</i> House	Chloroplast	MH173259	Chen <i>et al.</i> (2024)
<i>Ipomoea tabascana</i> J.A.McDonald & D.F.Austin	Chloroplast	MH173260	Chen <i>et al.</i> (2024)
<i>Ipomoea ternifolia</i> Cav.	Chloroplast	KF242494	Eserman <i>et al.</i> (2014)
<i>Ipomoea tricolor</i> Cav.	Chloroplast	KF242495	Eserman <i>et al.</i> (2014)
<i>Ipomoea trifida</i> (Kunth) G.Don	Chloroplast	KF242496	Eserman <i>et al.</i> (2014)
<i>Ipomoea trifida</i> (Kunth) G.Don	Chloroplast	KF242476	Eserman <i>et al.</i> (2014)
<i>Ipomoea trifida</i> (Kunth) G.Don	Chloroplast	KF242476.1, MH173261	Pan <i>et al.</i> (2023), Chen <i>et al.</i> (2024)
<i>Ipomoea trifida</i> (Kunth) G.Don	Nuclear	PRJNA428214	Wu <i>et al.</i> (2018)
<i>Ipomoea triloba</i> L.	Chloroplast	MH173262	Chen <i>et al.</i> (2024)
<i>Ipomoea triloba</i> L.	Nuclear	PRJNA428241	Wu <i>et al.</i> (2018)
<i>Merremia hederacea</i> (Burm. f.) Hall. f.	Mitochondrial	MZ240731	Lin <i>et al.</i> (2022)
<i>Merremia hungaiensis</i> (Lingelsh. & Borza) R.C.Fang	Chloroplast	OR771020	Chen <i>et al.</i> (2024)
<i>Operculina macrocarpa</i> (L.) Urb.	Chloroplast	KF242502	Eserman <i>et al.</i> (2014)
<i>Poranopsis discifera</i> (C.K.Schneid.) Staples	Chloroplast	OR771022	Chen <i>et al.</i> (2024)
<i>Stictocardia macalusoii</i> (Mattei) Verdc.	Chloroplast	KF242503	Eserman <i>et al.</i> (2014)
<i>Tridynamia sinensis</i> (Hemsl.) Staples	Chloroplast	OR771023	Chen <i>et al.</i> (2024)
<i>Turbina corymbosa</i> (L.) Raf.	Chloroplast	KF242504	Eserman <i>et al.</i> (2014)

Supplementary Material S3: References for Publications of Genomic Accessions

- ANDERSON B.M., KRAUSE K. & G. PETERSEN 2021. Mitochondrial genomes of two parasitic *Cuscuta* species lack clear evidence of horizontal gene transfer and retain unusually fragmented *ccmFC* genes. *BMC Genomics* 22: 816. <https://doi.org/10.1186/s12864-021-08105-z>
- BANERJEE A. & S. STEFANOVIĆ 2019. Caught in action: Fine-scale plastome evolution in the parasitic plants of *Cuscuta* section *Ceratophorae* (Convolvulaceae). *Plant Molecular Biology* 100(6): 621–634. <https://doi.org/10.1007/s11103-019-00884-0>
- BANERJEE A. & S. STEFANOVIĆ 2020. Reconstructing plastome evolution across the phylogenetic backbone of the parasitic plant genus *Cuscuta* (Convolvulaceae). *Botanical Journal of the Linnean Society*, 194(4): 423–438. <https://doi.org/10.1093/botlinnean/boaa056>
- CHEN L.Q., LI X., YAO X., LI D.Z., BARRETT C., DEPAMPHILIS C.W. & W.B. YU 2024. Variations and reduction of plastome are associated with the evolution of parasitism in Convolvulaceae. *Plant Molecular Biology*, 114(3): 40. <https://doi.org/10.1007/s11103-024-01440-1>
- ESERMAN L.A., TILEY G.P., JARRET R.L., LEEBENS-MACK J.H. & R.E. MILLER 2014. Phylogenetics and diversification of morning glories (tribe *Ipomoeae*, Convolvulaceae) based on whole plastome sequences. *American Journal of Botany*, 101(1): 92–103. <https://doi.org/10.3732/ajb.1300207>
- FU W., LI L., LONG L., WEN X., CHEN F., LI S. & Y. ZOU 2024. The complete chloroplast genome sequence of *Calystegia hederacea* Wall. In Roxb. 1824 (Convolvulaceae) in Enshi, Hubei. *Mitochondrial DNA Part B*, 9(5): 683–686. <https://doi.org/10.1080/23802359.2024.2336703>
- FUNK H.T., BERG S., KRUPINSKA K, MAIER, U.G. & K. KRAUSER 2007. Complete DNA sequences of the plastid genomes of two parasitic flowering plant species, *Cuscuta reflexa* and *Cuscuta gronovii*. *BMC Plant Biology*, 7: 45. <https://doi.org/10.1186/1471-2229-7-45>
- HOSHINO A., JAYAKUMAR V., NITASAKA E., TOYODA A., NOGUCHI H., ITOH T., SHIN-I.T., MINAKUCHI Y., KODA Y., NAGANO A.J., YASUGI M., HONJO M.N., KUDOH H., SEKI M., KAMIYA A., SHIRAKI T., CARNINCI P., ASAMIZU E., NISHIDE H. & Y. SAKAKIBARA 2016. Genome sequence and analysis of the Japanese morning glory *Ipomoea nil*. *Nature Communications*, 7(1): 13295. <https://doi.org/10.1038/ncomms13295>
- LIN Y., LI P., ZHANG Y., AKHTER D., PAN R., FU Z., HUANG M., LI X. & Y. FENG 2022. Unprecedented organelle genomic variations in morning glories reveal independent evolutionary scenarios of parasitic plants and the diversification of plant mitochondrial complexes. *BMC Biology*, 20(1): 49. <https://doi.org/10.1186/s12915-022-01250-1>
- MCNEAL J.R., KUEHL J.V., BOORE J.L. & C.W. DEPAMPHILIS 2007. Complete plastid genome sequences suggest strong selection for retention of photosynthetic genes in the parasitic plant genus *Cuscuta*. *BMC Plant Biology*, 7(1): 57. <https://doi.org/10.1186/1471-2229-7-57>
- NESBITT M. 2014. *Use of herbarium specimens in ethnobotany*. Curating biocultural collections. Royal Botanic Gardens, Kew. pp.313-28.
- PAN H., ZAGORCHEV L., CHEN L., TAO Y., CAI C., JIANG M., SUN Z. & J. LI 2023. Complete chloroplast genomes of five *Cuscuta* species and their evolutionary significance in the *Cuscuta* genus. *BMC Genomics* 24(1): 310. <https://doi.org/10.1186/s12864-023-09427-w>
- PARK I., SONG J.H., YANG S., KIM W.J., CHOI G. & B.C. MOON 2019. *Cuscuta* species identification based on the morphology of reproductive organs and complete chloroplast genome sequences. *International Journal of Molecular Sciences*, 20(11): 2726. <https://doi.org/10.3390/ijms20112726>
- PARK I., YANG S., KIM W.J., NOH P., LEE H.O. & B.C. MOON 2018. The complete chloroplast genome of *Cuscuta pentagona* Engelm. *Mitochondrial DNA B Resources*, 3(2): 523–524. <https://doi.org/10.1080/23802359.2018.1467229>
- SUDMOON R., KAEWDAUNGDEE S., HO H.X., LEE S.Y., TANEE T. & A. CHAVEERACH 2024. The chloroplast genome sequences of *Ipomoea alba* and *I. obscura* (Convolvulaceae): Genome comparison and phylogenetic analysis. *Scientific Reports*, 14(1): 14078. <https://doi.org/10.1038/s41598-024-64879-8>
- SUN J., DONG X., CAO Q., XU T., ZHU M., SUN J., DONG T., MA D., HAN Y. & Z. LI 2019. A systematic comparison of eight new plastome sequences from

Ipomoea L. *PeerJ* 7: e6563. <https://doi.org/10.7717/peerj.6563>

WU S., LAU K.H., CAO Q., HAMILTON J.P., SUN H., ZHOU C., ESERMAN L., GEMENET D.C., OLUKOLU B.A., WANG H., CRISOVAN E., GODDEN G.T., JIAO C., WANG X., KITAVI M., MANRIQUE-CARPINTERO N., VAILLANCOURT B., WIEGERT-RININGER K., YANG X. & Z. FEI 2018. Genome sequences of two diploid wild relatives of cultivated sweetpotato reveal targets for genetic improvement. *Nature Communications*, 9(1): 4580. <https://doi.org/10.1038/s41467-018-06983-8>

YANG J., MOEINZADEH M.H., KUHL H., HELMUTH J., XIAO P., HAAS S., LIU G., ZHENG J., SUN Z., FAN W., DENG G., WANG H., HU F., ZHAO S., FERNIE A.R., BOERNO S., TIMMERMANN B., ZHANG P. & M. VINGRON 2017. Haplotype- resolved sweet potato genome traces back its hexaploidization history. *Nature Plants*, 3(9): 696– 703. <https://doi.org/10.1038/s41477-017-0002-z>

YANG Z., NI Y., LIN Z., YANG L., CHEN G., NIJIATI N., HU Y. & X CHEN 2022. De novo assembly of the complete mitochondrial genome of sweet potato (*Ipomoea batatas* [L.] Lam) revealed the existence of homologous conformations generated by the repeat-mediated recombination. *BMC Plant Biology*, 22: 285. <https://doi.org/10.1186/s12870-022-03665-y>

Supplementary material S4: Suggested Molecular protocol for DNA extraction**DNA Extraction** (modified CTAB protocol, based on Doyle & Doyle, 1987)

CTAB Buffer:

- 10 ml of 1 M Tri-HCl
 - 28 ml of 5 M NaCl
 - 4 ml of 0.5 M EDTA
 - 2% CTAB (2g diluted in 100ml)
 - 2% PVP 40 (2g diluted in 100ml)
1. Label 1.5mL eppendorf tubes with sample number.
 2. Grind c. 20mg of dried leaf material using a pestle and mortar, or an automatic tissue lyser. Add a small amount of sand or glass beads to help grind fibrous tissue.
 3. Transfer all the powdered tissue into 1.5 mL eppendorf tubes using a clean spatula.
 4. Turn on heating block (or dry bath) at 65°C.
 5. In the fume cupboard: add, to each 1.5mL eppendorf tube:
 - 490µl CTAB buffer
 - 50µl Sarkosyl
 - 10µl Proteinase K
 6. Place all tubes in the heating block.
 7. Leave for 1 hour.
 8. Label additional 1.5 mL eppendorf tubes with sample number
 9. In the fume cupboard: add 500µl of SEVAC to each sample (1.5 mL eppendorf tube)
 10. Centrifuge for 3 minutes at 13,000rpm, to separate phases.
 11. Pipette off 250µl of the clear upper aqueous phase into a clean 1.5 mL eppendorf tube.
 12. Take care to avoid material at interface.
 13. **WARNING:** all the waste (liquid and solid) must be disposed of in the appropriate waste vessel (inside the fume cupboard)
 14. Purification
 15. Quantification: use a Spectrophotometer to quantify the DNA concentration in each sample.
 16. For blanking the measurement, always use the same substance that you used to elute the DNA in the final step of purification: buffer or water.
 17. Annotate values for DNA concentration and A260/A280 (purity), and curve shape.

Supplementary material S5: Markers and PCR conditions for Sanger sequencing
Recommended genetic markers for phylogenetic studies of Convolvulaceae

Genome	Region	Description	Marker	Primer Sequence
Nuclear	ITS	nuclear ribosomal internal transcribed spacer	ITS AB101	ACGAATTCATGGTCCGGTGAAGTGTTCCG
			ITS AB102	TAGAATTCCTCCGGTTCGCTCGCCGTTAC
Chloroplast	trnL-F	transfer RNA genes	trnL5 ^{UAAF} (TabC)	CGA AAT CGG TAGACG CTA CG
			trnF ^{GAA} (TabF)	ATT TGA ACTGGT GAC ACG AG
Chloroplast	matK	maturase K gene	matk390F	CGATCTATTCATTCAATATTTC
			matK1326R	TCTAGCACACGAAAGTCGAAGT
Chloroplast	rps16	rps16 gene	rps16x2F2	AAAGTGGGTTTTTATGATCC
			trnK(UUU)x1	TTAAAAGCCGAGTACTCTACC

PCR Conditions (suggested; based on Carine *et al.*, 2004; Shaw *et al.*, 2005)

Nuclear (ITS):

- **Initial denaturation:** 5 min at 94°C
- **Annealing:**

30 cycles of:

- 1 min at 94°C (denaturation)
- 3 min at 49°C (annealing)
- 1 min at 72°C (extension)
- **Final Extension:** 8 min at 72°C

Chloroplast markers (trnL-F, matK, rps16):

- **Initial denaturation:** 5 min at 80°C
- **Annealing:**

30 cycles of:

- 1 min at 95°C (denaturation)
- 1 min at 50°C (annealing)
- ramp: 0.38C/s until 65°C
- 4 min at 65°C (extension)
- **Final Extension:** 5 min at 65°C

Supplementary Table S6: Traditional uses of some Convolvulaceae species.

Species	Use	Part used	Region	Reference
<i>Argyreia nervosa</i> (Burm.f.) Bojer	Medicinal	Seeds	India	Padhi <i>et al.</i> (2013)
<i>Bonamia ferruginea</i> (Choisy) Hallier f.	Medicinal	Leaves	Brasil	Rodrigues (2006), Paes and Mendonça (2008)
<i>Cuscuta spp.</i>	Dye	Stem	Mexico	Wallert (1997)
<i>Decalobanthus mammosus</i> (Lour.) A.R.Simões & Staples	Food	Root	India	Staples (2022)
<i>Distimake tuberosus</i> (L.) A.R.Simões & Staples	Medicinal	Root	Mexico, Guinea	Urbina (1906), Mwangi Mwangi <i>et al.</i> (2022)
<i>Ipomoea aquatica</i> Forssk.	Food/ Medicinal	Leaves	Tropical Asia	Austin (2007), Malakar Paesand and Choudhury (2015)
<i>Ipomoea asarifolia</i> (Desr.) Roem. & Schult.	Fodder	Leaves	Pakistan	Ekenyem (2006)
<i>Ipomoea batatas</i> (L.) Lam.	Food	Root	Worldwide	Islam (2024)
<i>Ipomoea bracteata</i> Cav.	Food	Root	Mexico	Piedra-Malagón <i>et al.</i> , 2022
<i>Ipomoea capillacea</i> (Kunth) G.Don	Food	Root	Mexico	Sandoval-Ortega <i>et al.</i> (2023)
<i>Ipomoea caudata</i> Fernald	Fishing	Root	Mexico	Urbina (1906)
<i>Ipomoea corymbosa</i> (L.) Roth	Medicinal	Seeds	Mexico	Schultes & Hofmann (1973)
<i>Ipomoea dumosa</i> (Benth.) L.O. Williams	Food	Leaves	Mexico	Piedra-Malagón <i>et al.</i> (2022)
<i>Ipomoea jicama</i> Brandegees	Food	Root	Mexico	Urbina (1906)
<i>Ipomoea longituba</i> Hallier f.	Food	Root	Kenya	Kabuye (1986)
<i>Ipomoea mauritiana</i> Jacq.	Medicinal	Roots	India	Chandira and Jayakar (2010), Jain <i>et al.</i> (2011)
<i>Ipomoea oenotherae</i> (Vatke) Hallier f.	Food	Roots	Kenya	Kabuye (1986)
<i>Ipomoea purga</i> (Wender.) Hayne	Medicinal	Root	Mexico	Urbina (1906) Linajes <i>et al.</i> (1994)
<i>Ipomoea sinensis</i> (Desr.) Choisy	Food	Root	Kenya	Kabuye (1986)
<i>Ipomoea stans</i> Cav.	Medicinal	Root	Mexico	Díaz (1976)

Supplementary Material S7: References for Traditional Uses of Convolvulaceae Species.

- CHANDIRA M. & B. JAYAKAR 2010. Formulation and evaluation of herbal tablets containing *Ipomoea digitata* Linn. extract. *International Journal of Pharmaceutical Sciences Review and Research* 3(1): 022.
- DÍAZ J.L. 1976. Uso de las Plantas Medicinales de México. *Monografías Científicas II*. IMEPLAN, México D.F., México, pp. 31, 56–57, 67, 69, 118.
- EKENYEM B.U. 2006. An assessment of *Ipomoea asarifolia* leaf meal as feed ingredient in grower pig diet. *Pakistan Journal of Nutrition* 5(1): 39–42. <https://doi.org/10.3923/pjn.2006.39.42>
- ISLAM S. 2024. Super food of the next century? An intensive review on their potential as a sustainable and versatile food source for future generations. *CyTA - Journal of Food* 22 (1): 2397553. <https://doi.org/10.1080/19476337.2024.2397553>
- JAIN V., VERMA S.K. & S.S. KATEWA 2011. Therapeutic validation of *Ipomoea digitata* tuber (*Ksheervidari*) for its effects on cardio-vascular risk parameters. *Indian Journal of Traditional Knowledge* 10(4): 617–623.
- KABUYE C.H.S. 1986. Edible roots from wild plants in arid and semi-arid Kenya. *Journal of Arid Environments* 11(1): 65–74. [https://doi.org/10.1016/S0140-1963\(18\)31310-7](https://doi.org/10.1016/S0140-1963(18)31310-7)
- LINAJES A., RICO-GRAY V. & G. CARRIÓN 1994. Traditional production system of the root of jalapa, *Ipomoea purga* (Convolvulaceae), in central Veracruz, Mexico. *Economic Botany* 48(1): 84–89. <http://www.jstor.org/stable/4255576>
- MEIRA M., SILVA E., DAVID J. & J. DAVID 2012. Review of the genus *Ipomoea*: Traditional uses, chemistry and biological activities. *Revista Brasileira de Farmacognosia* 22: 682–713. <https://doi.org/10.1590/S0102-695X2012005000025>
- MWANGA MWANGA I.J.C., SOSEF M.S.M. & A.R.G. SIMÕES 2022. *Flore d'Afrique centrale. Convolvulaceae*. Jardin botanique de Meise, Meise.
- PADHI M., MAHAPATRA S., PANDA J. & N.K. MISHRA 2013. Traditional uses and phytopharmacological aspects of *Argyreia nervosa*. *Journal of Advanced Pharmaceutical Research* 4(1): 23–32.
- PAES L.S. & M.S. MENDONÇA 2008. Morphoanatomical aspects of *Bonamia ferruginea* (Choisy) Hallier f. (Convolvulaceae). *Revista Brasileira de Plantas Mediciniais* 10(4): 76–82.
- PIEDRA-MALAGÓN E.M., SOSA V., ANGULO D.F. & M.H. DÍAZ-TORIBIO 2022. Edible native plants of the Gulf of Mexico Province. *Biodiversity Data Journal* 10: e80565. <https://doi.org/10.3897/BDJ.10.e80565>
- SANDOVAL-ORTEGA M.H., LOERA-ÁVILA E.E., MARTÍNEZ-CALDERÓN V.M. & S.G. ZUMAYAMENDOZA 2023. Plantas silvestres comestibles del estado de Aguascalientes, México, sus formas de consumo y comercialización. *Polibotánica* 55: 213–230. <https://doi.org/10.18387/polibotanica.55.14>
- STAPLES G. 2022. A synoptic revision of the golden glories, genus *Decalobanthus* (Convolvulaceae). *Blumea - Biodiversity, Evolution and Biogeography of Plants* 67(1): 37–70.
- URBINA M. 1906. *Raíces comestibles entre los antiguos Mexicanos*. Imprento del Museo Nacional, Mexico.
- WALLERT A. 1997. The analysis of dyestuffs on historical textiles from Mexico. *The Unbroken Thread: Conserving the Textile Traditions of Oaxaca*: 57–85.